

UC Davis

UC Davis Previously Published Works

Title

Biostimulants in Plant Science: A Global Perspective.

Permalink

<https://escholarship.org/uc/item/2892271g>

Authors

Yakhin, Oleg I
Lubyanov, Aleksandr A
Yakhin, Ildus A
et al.

Publication Date

2016

DOI

10.3389/fpls.2016.02049

Peer reviewed



Biostimulants in Plant Science: A Global Perspective

Oleg I. Yakhin^{1,2*}, Aleksandr A. Lubyantsov², Ildus A. Yakhin² and Patrick H. Brown³

¹ Institute of Biochemistry and Genetics, Ufa Scientific Center, Russian Academy of Sciences, Ufa, Russia, ² R&D Company Eco Priroda, Ulkundy, Russia, ³ Department of Plant Sciences, University of California, Davis, Davis, CA, USA

This review presents a comprehensive and systematic study of the field of plant biostimulants and considers the fundamental and innovative principles underlying this technology. The elucidation of the biological basis of biostimulant function is a prerequisite for the development of science-based biostimulant industry and sound regulations governing these compounds. The task of defining the biological basis of biostimulants as a class of compounds, however, is made more complex by the diverse sources of biostimulants present in the market, which include bacteria, fungi, seaweeds, higher plants, animals and humate-containing raw materials, and the wide diversity of industrial processes utilized in their preparation. To distinguish biostimulants from the existing legislative product categories we propose the following definition of a biostimulant as “a formulated product of biological origin that improves plant productivity as a consequence of the novel or emergent properties of the complex of constituents, and not as a sole consequence of the presence of known essential plant nutrients, plant growth regulators, or plant protective compounds.” The definition provided here is important as it emphasizes the principle that biological function can be positively modulated through application of molecules, or mixtures of molecules, for which an explicit mode of action has not been defined. Given the difficulty in determining a “mode of action” for a biostimulant, and recognizing the need for the market in biostimulants to attain legitimacy, we suggest that the focus of biostimulant research and validation should be upon proof of efficacy and safety and the determination of a broad mechanism of action, without a requirement for the determination of a specific mode of action. While there is a clear commercial imperative to rationalize biostimulants as a discrete class of products, there is also a compelling biological case for the science-based development of, and experimentation with biostimulants in the expectation that this may lead to the identification of novel biological molecules and phenomenon, pathways and processes, that would not have been discovered if the category of biostimulants did not exist, or was not considered legitimate.

Keywords: biostimulants, mode of action, definition, classification, regulation, concepts, methodology, emergent properties

INTRODUCTION

The regulation of plant growth and the development and alleviation of the negative effects of environmental stresses during ontogenesis, are important factors determining the productivity of cultivated plants. While it is well recognized that biotic and abiotic stress prevents essentially all crop systems from achieving their yield potential, current understanding of the mechanisms

OPEN ACCESS

Edited by:

Stefania De Pascale,
University of Naples Federico II, Italy

Reviewed by:

Yuksel Tuzel,
Ege University Faculty of Agriculture,
Turkey

Patrick Du Jardin,
Gembloux Agro-Bio Tech - University
of Liège, Belgium

*Correspondence:

Oleg I. Yakhin
yakhin@anrb.ru

Specialty section:

This article was submitted to
Crop Science and Horticulture,
a section of the journal
Frontiers in Plant Science

Received: 27 October 2016

Accepted: 21 December 2016

Published: 26 January 2017

Citation:

Yakhin OI, Lubyantsov AA, Yakhin IA
and Brown PH (2017) Biostimulants in
Plant Science: A Global Perspective.
Front. Plant Sci. 7:2049.
doi: 10.3389/fpls.2016.02049

involved, and the strategies to mitigate these effects are limited. Abiotic stresses may be prevented by optimizing plant growth conditions and through provision of water and nutrients and plant growth regulators (PGRs—auxins, cytokinins, gibberellins, strigolactones, brassinosteroids). In addition to these traditional approaches, biostimulants are increasingly being integrated into production systems with the goal of modifying physiological processes in plants to optimize productivity. Plant biostimulants based on natural materials have received considerable attention by both the scientific community and commercial enterprises especially in the last two and a half decades (Crouch and van Staden, 1993a; Herve, 1994; Zhang and Schmidt, 1999; Maini, 2006; Khan et al., 2009; Apone et al., 2010; Craigie, 2011; Sharma et al., 2014; Brown and Saa, 2015; Du Jardin, 2015; Yakhin et al., 2016a). Biostimulants offer a potentially novel approach for the regulation/modification of physiological processes in plants to stimulate growth, to mitigate stress-induced limitations, and to increase yield. In the following review, we do not attempt to discern if the effects of biostimulants on plant productivity is a direct response of plants or soils to the biostimulant application or an indirect response of the biostimulant on the soil and plant microbiome with subsequent effects on plant productivity. Ultimately discerning if biostimulant effects are direct or microbially mediated will be critical to the development of this technology. The general goals of the current review are to provide a comprehensive analysis of the current situation in the field of biostimulants and to develop a science-based theoretical foundation for the conceptualization, classification, and practical application of these materials. A focus of this review is to understand and define the appropriate place of biostimulants among other agricultural products such as plant protection compounds and fertilizers, and to consider the unique attributes of complex, multi-component biostimulants. The structure of the review is based on the consideration of biostimulants in terms of their action on different regulatory and functional systems of plants (signaling, metabolism, uptake, and transport mechanisms, etc.) using both conceptual and methodological approaches. The overarching objective of the work is to highlight innovative concepts and to establish a scientific framework for future development of biostimulant science.

GENERAL CONCEPTS AND METHODOLOGY

To understand the development of biostimulant science, several seminal publications warrant discussion. To our knowledge, the first discussion of “biogenic stimulant” theory can be attributed to Prof. V.P. Filatov and was started in 1933 in the USSR (Filatov, 1944, 1951a,b; Gordon, 1947; Sukhovkhorov, 1967). Filatov proposed that biological materials derived from various organisms, including plants, that have been exposed to stressors could affect metabolic and energetic processes in humans, animals, and plants (Table 1). Blagoveshchensky (1945, 1955, 1956) further developed these ideas with specific reference to their application for plants, considering biogenic stimulants as “organic acids with stimulating effects due to their

dibasic properties which can enhance the enzymatic activity in plants.” Filatov’s concept (1951b), was, however, not limited to these compounds alone (Filatov, 1951b). Herve’s (1994) pioneering review provides the first real conceptual approach to biostimulants. Herve suggests the development of novel “bio-rational products” should proceed on the basis of a systemic approach founded in chemical synthesis, biochemistry, and biotechnology as applied to real plant physiological, agricultural, and ecological constraints. He suggests these products should function at low doses, be ecologically benign and have reproducible benefits in agricultural plant cultivation. Zhang and Schmidt (1999) emphasized the need for comprehensive and empirical analysis of these products with particular emphasis on hormonal and antioxidant systems as the basis for many important benefits of biostimulants. They discuss the concept of biostimulants as “pre-stress conditioners,” their effects being manifested in improved photosynthetic efficiency, reduction of spread and intensity of some diseases and in better yields. Basak (2008) initiated the systematic discussion on biostimulants and created the conceptual preconditions for the formation of present biostimulant science while Du Jardin (2012, 2015) provided the first in-depth analysis of plant biostimulant science with an emphasis on biostimulant systematization and categorization on the basis of biochemical and physiological function and mode of action and origin. Du Jardin’s (2015) analysis and categorization was influential in informing the development of subsequent legislation and regulation in the European Union.

The study and development of biostimulants has been approached utilizing a wide range of methodological approaches including chemical and non-chemical characterization of composition (Crouch and van Staden, 1993b; Yakhin et al., 2005; Parrado et al., 2008; Sharma et al., 2012a,b; Ertani et al., 2013a,b; Aremu et al., 2015a,b), plant growth and yield studies (Khan et al., 2009; Kunicki et al., 2010; Paradiković et al., 2011; Zodape et al., 2011; Yakhin et al., 2012, 2016b; Chbani et al., 2013; Kurepin et al., 2014; Colla et al., 2015; Saa et al., 2015; Tandon and Dubey, 2015; Tian et al., 2015), application of the so-called *-omics* strategies with variations, including microarray and physiological analysis (Jannin et al., 2012, 2013), transcriptome (Wilson et al., 2015; Goñi et al., 2016), genomic (Santaniello et al., 2013), phenomic and molecular (Petrozza et al., 2014), proteomic (Martínez-Esteso et al., 2016), chemical and metabolomic (Ertani et al., 2014). Ultimately, the integrative synthesis of results from multiple methodologies, particularly when integrated with the most relevant—*omic* technology, “agronomics,” will be required if the science and legitimacy of plant biostimulants is to advance.

Several significant scientific meetings in the field of biostimulants have been held over the past ten years and have contributed greatly to our understanding of conceptual and methodological development of the biostimulant theory: “Biostimolanti in agricoltura” (Italy, 2006), “Biostimulators in Modern Agriculture” (Poland, 2008), “Biostimulants and Plant Growth” (Belgium, 2014), among others. Of particular significance were the first (France, 2012) and the second (Italy, 2015) World Congresses on the “Use of Biostimulants in Agriculture” which were valuable in highlighting the development of novel concepts and methodology as applied

TABLE 1 | Terminology in the biostimulant field: Evolution and diversity of concepts*.

Terms, variants, and synonyms of the term of "biostimulant"	Original definitions and hypotheses Translation from Russian publications into English is verbatim (word for word, literatim)	References
Biogenic stimulators	"Every living tissue (human, animal and plants), when exposed to unfavorable, but non-lethal conditions, undergoes biochemical restructuring with the formation in it of special substances which are biogenic stimulators of non-specific nature, stimulating the life reactions of the organism, in which they introduced in, one way or another."	Filatov, 1951a
Biogenic stimulants	"1. Organisms, either animal or plant, when exposed to such environmental factors that complicate their lives, are subjected to biochemical restructuring. Consequently, there are formed substances that stimulate biochemical processes in these tissues. These substances which help the tissues to preserve life under adverse conditions, are named stimulants of biological origin (biogenic stimulators)." "2. Biogenic stimulators, injected one way or another in any organism activates vital processes in it. By strengthening metabolism, thus they increase physiological functions of the organism. In this manner biogenic stimulants increase the organism's resistance to pathogenic factors and enhance its regenerative and absorbable properties, which facilitates recovery." "3. Biogenic stimulators emerge as a result of biochemical restructuring, and in whole living organisms subjected to non-lethal but unfavorable external or internal environmental conditions." "4. Environmental factors that cause the emerging of biogenic stimulants in the organism or in tissues separated from it, can be diverse." "5. The emergence of biogenic stimulants under the influence of unfavourable factors of the environment is a common law for all wildlife. Biogenic stimulators are formed wherever there is a adaptation to new conditions of existence and the struggle for life." "6. Biogenic stimulators accumulate in tissues and organisms when exposed to such external and internal factors that lead to the disruption of their normal metabolism, and are chemically products of disturbed metabolism." "7. Biogenic stimulants act on the whole organism. This explains the breadth of the range of their action on the organism." "8. The action of biogenic stimulators is expressed in changing of metabolic and energetic processes of organism."	Filatov, 1951b
Biogenic stimulants	"Substances which are produced in living tissues when using the method of Filatov following a series of disturbances of normal metabolism for the organism (according to Filatov - resistance factors), that have a stimulating effect on various processes in the organism." [sic] "Biogenic stimulators can not substitute for fertilizer."	Blagoveshchensky, 1956
Organic Biostimulant	"These compounds increase plant growth and vigor through increased efficiency of nutrient and water uptake. Definitions for biostimulants vary greatly and there are still some arguments surrounding these compounds. However, they are defined as non-fertilizer products which have a beneficial effect on plant growth. Many of these biostimulant materials are natural products that contain no added chemicals or synthetic plant growth regulators."	Russo and Berlyn, 1991
Biostimulators	"Materials of little or no fertilizer value that accelerate plant growth, usually when used at low concentrations."	Goatley and Schmidt, 1991
Biostimulants	"Plant hormone-containing substances that can stimulate growth when exogenously applied."	Schmidt, 1992
Allelopathic Preparation	"Multi-component balanced systems of biologically active substances of metabolic origin on the basis of plant raw materials with a broad spectrum of biological activity."	Naumov et al., 1993
Biostimulants	A subgroup of plant growth regulators but are quite different from nutritional additives. ... It is proposed to limit the use of the term biostimulant to products aimed at improving yield through various metabolistics pathways.	Herve, 1994
Biostimulants	"Products that are nonnutritive promoters of growth. Growth can be promoted by stimulating nutrient uptake, chelating nutrients, providing plant growth hormones, or enhancing plant hormonal activity. Biostimulants that contain plant growth hormones can be produced synthetically or obtained from natural plant extracts."	Elliott and Prevatte, 1996
Biostimulant	"Materials that, in minute quantities, promote plant growth."	Zhang and Schmidt, 1999
Biostimulant	"An ambiguous term used to encompass non-nutritional growth-promoting substances such as microbes, plant growth hormones, soil conditioners and microbe energy sources."	McCarty, 2001
Plant Strengtheners	"Products intended to protect plants against harmful organisms by stimulating defence mechanisms in the plant or by competing with harmful organisms for space and nutrients in the phyllosphere or rhizosphere."	(Anonymous 2001) quoted by Sharma K. et al., 2012
Biostimulant (Positive Plant Growth Regulator), (Metabolic Enhancer)	An organic material that, when applied in small quantities, enhances plant growth and development such that the response cannot be attributed to application of traditional plant nutrients. ... If applied before stress occurs, biostimulants can help plants tolerate stress.	James Beard from Schmidt et al., 2003
Biostimulants	"Natural or synthetic products of either mineral or organic composition that by their mode of action positively contribute to crop nutrition and the development of healthy plants."	(S.D. Hankins, personal communication) Dixon and Walsh, 2004

(Continued)

TABLE 1 | Continued

Terms, variants, and synonyms of the term of "biostimulant"	Original definitions and hypotheses Translation from Russian publications into English is verbatim (word for word, literatim)	References
Metabolic Enhancer	"Non-mineral substances that, when exogenously applied in very small quantities, stimulate the metabolic activities in plants."	Doak et al., 2005
Biostimolanti - Biostimulants	"Products that brings to other fertilizer and/or to the soil and/or to the plant, substances that favour or regulate the absorption of the nutrients or correct some physiological anomalies." "It must be remind that the biostimulant activity does not have to derive from the addition of phytohormones (Attached 6, codicil 4.1). In other words, a fertilizer with "biostimulant activity" must contribute positively to the improvement of the nutrition and the development of the plants, independently from the presence of the nutrients, with the exclusion of the phytohormones, whose presence is prohibited, and of the products with declared and specific phytosanitary function."	Ciavatta and Cavani, 2006
Biostimulants	"This term commonly identifies formulations supporting the plant in the improvement of its performances without using synthesis hormones."	Tagliavini and Kubiskin, 2006
Organic Biostimulant	"Other than the direct influence on the nitrogen balance in plants, Siapton acts also indirectly improving the activity of some enzyme systems and regulating some plant growth regulators (PGR) functions and biochemical processes. Moreover, Siapton makes easier the absorption and the transport of the nutritional macro- and micro-elements. These statements allow to define the product as a real "organic biostimulant" and natural nutritional equilibrators for plants, more than a simple foliar organic fertilizer."	Maini, 2006
Biostimulants	"Product generally of organic nature which increase the soil microbial activity and/or plant growth."	Nardi et al., 2006
Biostimulants	"Single compounds or mixtures of compounds which mitigate the effects caused by abiotic stress agents, through the induction of innate plant defense responses."	Apone et al., 2006
Biostimulant Plant Growth Promoters Growth-Promoting PGRs	"Organic or hormone-containing compounds derived from natural products which can stimulate plant growth and development without causing known harm to the environment."	Huang, 2007
Biostimulators	"Compounds of biological origin and should act by increasing natural capabilities of plants to cope with stresses. Biostimulators do not act neither as nutrients nor affect directly the stress factors making them less harmful for plants."	Gawrońska, 2008
Biostimulators	"Agents which at very low concentrations improve the basic biochemical processes in plants and soil, and thereby improve the growth and development of plants, and increase their resistance to stress. Biostimulators are not a substitute for fertilizers, manure or other sources of mineral nutrients."	Anonymous, cited by Basak, 2008
Biostimulators	"Innovative natural stimulators of plant growth and development, obtained from biological raw materials, and having a hormone- or fertilizer-like mode of action."	Anonymous, cited by Basak, 2008
Biofertiliser/Biostimulant	"Are organic products composed of peptides, amino acids, polysaccharides, peptides, humic acids, and/or phytohormones, etc. for immediate uptake and availability within the plant. Their absorption does not depend on the photosynthetic activity as they are directly absorbed by the plant, resulting in lower energy consumption. The aim of these products is not to supply nutrition, but rather to favour and stimulate the metabolism of the plant, decrease plant stress, etc."	Parrado et al., 2008
Organic Biostimulants	"Non-nutrient containing products which have beneficial effect on plant growth. Such products consist of humic acids, marine algae (sea weed) extracts, non-hormonal plant metabolites and vitamins."	Kumar and Shivay, 2008
Phytostimulator	"Microorganism with the ability to produce or change the concentration of growth regulators such as indole acetic acid, gibberellic acid, cytokinins and ethylene."	Martínez-Viveros et al., 2010
Agricultural Biostimulants	"Include diverse formulations of compounds, substances and other products that are applied to plants or soils to regulate and enhance the crop's physiological processes, thus making them more efficient. Biostimulants act on plant physiology through different pathways than nutrients to improve crop vigour, yields, quality and post-harvest shelf life/conservation."	European Biostimulants Industry Council EBIC, 2011b
Biostimulators	"They mean inorganic and organic substances or its mixtures positively affecting plant development or other physiological processes in plants. One of the requirements for biostimulators is that they pose no risk for human, animal, or natural environment due to its application."	Smoleń S, 2012
Biostimulants	"Materials that are neither a fertilizer nor a pesticide, but when applied to a plant will enhance their health, growth, and protection."	Banks and Percival, 2012
Plant Biostimulants	"Substances and materials, with the exception of nutrients and pesticides, which, when applied to plants, seeds or growing substrates in specific formulations, have the capacity to modify physiological processes of plants in a way that provides potential benefits to growth, development and/or stress response."	Du Jardin, 2012
Antitranspirant	"Indicates the overall effect on the plant, the chemical compounds and underlying mechanisms are very diverse. Some of the compounds have physical effects at the surface and/or within the plant organs, others are regulators of the leaves openings diffusing water vapor, called stomata."	Du Jardin, 2012
Metabolic Antitranspirants	"Stomatal regulators, acting on the complex hormonal control of the highly specialized cells bordering the stomatal pore (guard cells)."	Du Jardin, 2012

(Continued)

TABLE 1 | Continued

Terms, variants, and synonyms of the term of “biostimulant”	Original definitions and hypotheses Translation from Russian publications into English is verbatim (word for word, literatim)	References
Plant Biostimulants	“Contain substance(s) and/or micro-organisms whose function when applied to plants or the rhizosphere is to stimulate natural processes to enhance/benefit nutrient uptake, nutrient efficiency, tolerance to abiotic stress, and crop quality. Biostimulants have no direct action against pests, and therefore do not fall within the regulatory framework of pesticides.”	European Biostimulants Industry Council EBIC, 2012
Biostimulant	“Material that, when applied to a plant, seed, soil or growing media - in conjunction with established fertilization plans, enhances the plant's nutrient use efficiency, or provides other direct or indirect benefits to plant development or stress response.”	Beaudreau, 2013, Biostimulant Coalition
Biostimulants	“Compounds that produce non-nutritional plant growth responses and reduce stress by enhancing stress tolerance. This is in contrast to fertilizers, which produce a nutritional response. Many important benefits of biostimulants are based on their ability to influence hormonal activity.” “Compounds in biostimulants can alter the hormonal status of a plant and exert large influences over its growth and health.”	Daniels, 2013
Plant Strengtheners (Biostimulant)	A class of “borderline” products used in agriculture in some member states ... which act only on plant strength without direct effects against pests and have no main nutritional activity, ... enhance the resistance of plants to harmful organisms and protect plants against non-parasitic impairments.”	Torre et al., 2013
Biostimulant	“Products which, alone or mixed with other fertilizers, contribute to improve plant growth by exploiting different physiological processes.”	Migliore et al., 2013
Bio-Stimulatory Bio-stimulatory Agent	“The term “bio-stimulatory” means according to the invention, if not otherwise specified, an activity or efficacy which stimulates, increases or improves many different processes in the plant or plant parts, such as improved generation of growth promoting substances like sugars and amino acids, improved adequate supply of cells with available nutrients and growth regulators, enhanced cell metabolism, improved cell decontamination, enhanced immune defense, promotion of growth and yield, induction of systemic acquired resistance (SAR), inhibition of growth and yield of competing plants (allelopathy). The bio-stimulatory activity can be caused by agents, plant extracts and compositions including metabolic compounds synthesized by the plant to be protected after induction of their synthesis by said bio-stimulatory agent. A “bio-stimulatory agent” according to the invention is a biological plant protecting agent as specified above, which shows the above-specified bio-stimulatory properties in a plant treated with this agent <i>in vitro</i> and/or <i>in vivo</i> .”	Pretorius, 2013
Biostimulant	“Is an organic material that, when applied in small quantities, enhances plant growth and development such that the response cannot be attributed to the application of traditional plant nutrients.”	Sharma et al., 2014
Biostimulant Microorganisms	“Both biocontrol microorganisms (BCMs) and plant growth-promoting microorganisms (PGPMs) can be defined as “biostimulant microorganisms,” able to foster plant growth and defence against pathogens throughout the crop life cycle, from seed germination to plant maturity.”	Sofo et al., 2014
Phytostimulators Biostimulators	“This category includes microorganisms that promote plant growth usually by hormonal action.”	Aguado-Santacruz et al., 2014
Plant Biostimulant	“Any substance or microorganism, in the form in which it is supplied to the user, applied to plants, seeds or the root environment with the intention to stimulate natural processes of plants benefiting nutrient use efficiency and/or tolerance to abiotic stress, regardless of its nutrients content, or any combination of such substances and/or microorganisms intended for this use.”	Traon et al., 2014
Biostimulants	“Are extracts obtained from organic raw materials containing bioactive compounds.”	Bulgari et al., 2015
Biostimulants	“Are materials, other than fertilizers, that promote plant growth when applied in small quantities. These environmental friendly and natural substances promote vegetative growth, mineral nutrient uptake and tolerance of plants to abiotic stresses.”	Chojnacka et al., 2015
Biostimulant	“Any substance or microorganism applied to plants with the aim to enhance nutrition efficiency, abiotic stress tolerance and/or crop quality traits, regardless of its nutrients content. By extension, plant biostimulants also designate commercial products containing mixtures of such substances and/or microorganisms.”	Du Jardin, 2015
Biostimulants	“Are substances or materials, with the exception of nutrients and pesticides, which, when applied to plants, seeds, or growing substrates in specific formulations, have the capacity to modify physiological processes in plants in a way that provides potential benefits to growth, development, or stress response.”	Halpern et al., 2015
Biostimulant	“Refers to a compound or composition that is neither a fertilizer nor pesticide, but which when applied to a plant will enhance the health and growth of a plant.”	Lovatt, 2015
Biostimulants	“Products mostly based on natural raw materials, used in the ultra-small and small doses for modification of physiological and biochemical plant processes with the aim of more complete realization of genetic potential of their productivity due to changes in hormonal status, activation of metabolic processes, increase of efficiency of nutrition, stimulation of growth, development and strengthening the ability to withstand to the negative effects of various stress factors.”	Yakhin et al., 2016a

*The definitions are provided as exact quotes from the primary sources without correction of spelling or grammar.

to biostimulants. While many of the following papers are not published in a peer-reviewed format, they do represent important advances in this field. Dumas et al. (2012), for example, proposed a multi-part approach to study biostimulants based on large-scale genomic approaches and high-throughput screening tests with genetically-modified reporter plants. Others suggested that biostimulant mode of action can be best determined using molecular microarray analysis to identify gene changes in transcript levels (Gates et al., 2012). This approach has the potential to reveal biostimulant activated signaling pathways involved in the stimulation of plant response. Microarray analysis is not, however, adequate and must be supplemented with carefully conducted field testing or high throughput plant phenotyping (Summerer et al., 2013). The complexity of known biostimulant response, the dependency of crop environment and the diversity of biostimulant products demands the application of novel statistical approaches not commonly used in agronomic research (Sleighter et al., 2015). The principle espoused by Sleighter et al. (2015) is based on the identification of a subset of molecular markers that represent the active ingredients in complex biostimulants and then to correlate these markers with observations of plant response. Chemical genomics that utilizes small molecules to perturb target protein function is a useful strategy for biostimulant discovery as it overcomes constraints imposed by traditional molecular approaches that often fail due to gene redundancy and loss-of-function lethality. Botta et al. (2015) proposed probing the function of biostimulants using an enantiomeric analysis of active compounds in the biostimulant coupled with a proteomic profiling approach. In contrast, Conan et al. (2015) proposed identification of the bioactive compounds responsible for the plant growth response by means of a metabolomic profiling of biostimulant products and analysis of their physiological effects through transcriptomic and metabolomic strategies. Such methodology allows the determination of metabolite pathways affected by biostimulants as well as providing insight into gene regulation. To integrate the diversity of methodologies available Santaniello et al. (2015) emphasizes the need to use bioinformatics strategies to analyse similarities and differences in procedures of ingredient extraction and biostimulant formulation in terms of molecular plant responses. This integrative concept can be used to derive new technologies and novel biostimulant products through the identification of new target genes, enzymes and metabolites.

While the development of robust, multi-faceted approach to the analysis of biostimulant composition and function will greatly aid in the development of this field, all advances must ultimately be interpreted in the context of plant response. The complexity of plant response to the environment is daunting and was elegantly highlighted by Krouk (2015) who demonstrated that root response to nitrogen in the environment is mediated by combinations of signaling molecules and nitrogen sources in a manner that cannot be predicted by exposure to single compounds provided individually (Krouk, 2016; Krouk et al., 2009, 2010, 2011). Inevitably, as our understanding of the molecular networks that control plant growth improves our ability to predict plant response to biostimulants under specific environmental conditions, will improve. Only through

a combination of methodologies will progress in biostimulant research be possible.

TERMINOLOGY AND DEFINITIONS

The development of plant biostimulant science, as well as the principles governing its legislation in the context of the existing legal frameworks of plant protection products and fertilizers, requires the development of a clear definition of term “biostimulant.” Currently, the term “biostimulant” is poorly defined and includes many products that have variously been described as biogenic stimulants, metabolic enhancers, plant strengtheners, positive plant growth regulators, elicitors, allelopathic preparation, plant conditioners, phytostimulators, biofertilisers, or biofertiliser/biostimulant (**Table 1**). One area of significant challenge is evoked in the question “are biostimulants PGRs?” Historically, biostimulants have been considered as a subgroup of growth regulators (Herve, 1994), as plant growth regulators (Huang, 2007), and as subgroup of bioregulators (Basak, 2008). “From a legal point of view, biostimulants can contain traces of natural plant hormones, but their biological action should not be ascribed to them, otherwise they should be registered as plant growth regulators” (Bulgari et al., 2015). Likewise, biostimulants cannot by definition be pesticides or fertilizers (Russo and Berlyn, 1991; Karnok, 2000; Hamza and Suggars, 2001; Banks and Percival, 2012; Du Jardin, 2012; Torre et al., 2013, 2016).

A concise and biologically meaningful definition of biostimulants has eluded researchers and regulators for many years. **Table 1** presents a chronological evolution of concept of the term biostimulant. While several of biostimulant definitions presented are useful in their breadth, many of them have significant limitations and are overly generic, while several do not exclude possible effects of nutrients contained within any putative biostimulant product. In practice, biostimulants may deliberately include nutrients for regulatory approval as fertilizers and on occasions the included nutrients or hormones may be responsible for the perceived agronomic benefit. Given the state of public mistrust of many “biostimulant” products, it is necessary to provide a definition of biostimulants that explicitly denies the use of this term for products that do not have biological efficacy or have efficacy only by virtue of the inclusion of known plant hormones or nutrients.

While the adoption of a definition of biostimulants for regulatory purposes is important, any definition of biostimulant should also be based on scientific principles. Several concepts have been proposed to define plant biostimulants. Basak (2008), proposed that biostimulants could be classified depending on the mode of action and the origin of the active ingredient while Bulgari et al. (2015), proposed that “biostimulants should be classified on the basis of their action in the plants or, on the physiological plant responses rather than on their composition.” Du Jardin (2015), however, has emphasized the importance of the final impact on plant productivity when he suggests that “any definition of biostimulants should focus on the agricultural functions of biostimulants, not on the nature of their constituents nor on their modes of actions.” The term

“plant productivity” is used here to describe any improvement in plant yield or quality or increased efficiency of production. These concepts reflect important differences in approaches to providing a definition of biostimulants as a discrete category of agricultural products. Thus, biostimulants could be defined by their demonstrated mode of action and origin, or solely by their demonstrated beneficial impact on plant productivity. The challenges in developing a definition are also complicated by the multi-component and largely undefined composition of many biostimulant products and the possibility that the activity of a biostimulant may not be explained by the presence of any individual constituent, but is a result of the interaction of many constituents in the product.

On this basis two approaches to the definition of complex biostimulants emerge. The first is based on the possibility that the biostimulant contains within it, previously unrecognized molecules that are the sole and discrete cause of the observed improvement in plant productivity. This concept emphasizes both the need for clear demonstration of plant productivity benefits and the unknown nature of the mode of action. Thus, a biostimulant could be defined as “a formulated product that improves plant productivity by a mechanism of action that is not the sole consequence of the presence of known essential plant nutrients, plant hormones, plant growth regulators or plant protective compounds.” By this definition, once the primary biological mechanism of biostimulant function has been identified it should henceforth, be subject to classification on the basis of that functional component.

The majority of biostimulants in use today are complex mixtures of chemicals derived from a biological process or extraction of biological materials. The complexity of these mixtures is often considered to be essential to the performance of the biostimulant, and biostimulants may have properties of the whole, that cannot be fully elucidated by knowing the characteristics of the separate components or their combinations. This theory of complexity or “emergence” was described by Mayr (1982), who argued that in many biological systems “the properties of the whole cannot be fully elucidated by knowing the characteristics of the separate components or their combinations.” “The term emergence describes the onset of novel properties that arise when a certain level of structural complexity is formed from components of lower complexity. In the last few decades, emergence has been discussed in a number of different research fields, such as cybernetics, theory of complexity, artificial intelligence, non-linear dynamics, information theory, and social systems organization” (Luisi, 2002). “Emergence” and “emergent properties” are thus closely related with the notion of the “systems biology” (Luisi, 2002; Johnson, 2006; Korosov, 2012; Lüttge, 2012; Bertolli et al., 2014). Emergence was described by Johnson (2006) as “unexpected behaviors that stem from interaction between the components of an application and their environment,” “there is, however, considerable disagreement about the nature of ‘emergent properties.’ Some include almost any unexpected properties exhibited by a complex system. Others refer to emergent properties when an application exhibits behaviors that cannot be identified through functional decomposition. In other words, the

system is more than the sum of its component parts” (Johnson, 2006).

Thus, a biostimulant could also be defined as “a formulated product of biological origin that improves plant productivity as a consequence of the emergent properties of its constituents.”

To our knowledge, however, there have been no clear demonstrations that any biostimulant exhibits truly emergent properties. This is not however a unique challenge and all “biological systems are extremely complex and have emergent properties that cannot explained, or even predicted, by studying their individual parts” (Van Regenmortel, 2004). Emergent properties have been demonstrated in the networks of biological signaling pathways (Bhalla and Iyengar, 1999); in system-level study of traditional Chinese medicine (Chen et al., 2014), and in microbial communities (Wintermute and Silver, 2010; Chiu et al., 2014). To adequately explain the biological complexity present in plants and their interactions with the environment, Lüttge (2012) and Bertolli et al. (2014) emphasize that classic reductionist biology/chemistry is indeed insufficient.

While the two theoretical definitions provided in this section share a requirement that the mode of action is unknown, they differ in the core assumption that biostimulant function is a consequence of the discrete components in the biostimulant or as a consequence of the “emergent” properties of the biostimulant as a whole. Each of these definitions is also incomplete in that it is certainly possible that a biostimulant may contain several molecules that act synergistically while not being truly “emergent,” and it is indeed possible and indeed likely, that even if a biostimulant is demonstrated to have emergent properties, that not all components of that biostimulant are required for that property to be expressed.

We propose, therefore, a definition of a biostimulant that integrates these two concepts. Thus, a biostimulant is defined here as:

“a formulated product of biological origin that improves plant productivity as a consequence of the novel, or emergent properties of the complex of constituents, and not as a sole consequence of the presence of known essential plant nutrients, plant growth regulators, or plant protective compounds.”

Consistent with this definition, the ultimate identification of a novel molecule within a biostimulant that is found to be wholly responsible for the biological function of that biostimulant, would necessitate the classification of the biostimulant according to the discovered function.

CLASSIFICATION

A review of the history of biostimulants and related products provides insight into the diversity of these products and the development of this field of study. The evolution of biostimulant classifications as described by various authors is presented in the **Table 2**. To the best of our knowledge, one of the first attempts to categorize biostimulants was provided by Filatov (1951b) when 4 groupings of biogenic stimulants were suggested. Karnok (2000) compiled a list of 59 materials presenting in

TABLE 2 | Proposed Biostimulant Categories.

Filatov, 1951b	Ikrina and Kolbin, 2004	Kauffman et al., 2007	Du Jardin, 2012	Calvo et al., 2014	Halpern et al., 2015	Du Jardin, 2015	Torre et al., 2016
1 Carboxylic fatty acids (oxalic acid and succinic acid)	Microorganisms (bacteria, fungi)	Humic substances	Humic substances	Microbial inoculants	Humic substances	Humic and fulvic acids	Humic substances
2 Carboxylic fatty hydroxy acids (malic and tartaric acids)	Plant materials (land, freshwater and marine)	Hormone containing products (seaweed extracts)	Complex organic materials	Humic acids	Protein hydrolysate and amino acid formulations	Protein hydrolysates and other N-containing compounds	Seaweed extracts
3 Unsaturated fatty acids, aromatic and phenolic acids (cinnamic and hydroxycinnamic acids, coumarin)	Sea shellfish, animals, bees	Amino acid containing products	Beneficial chemical elements	Fulvic acids	Seaweed extract	Seaweed extracts and botanicals	Hydrolyzed proteins and amino acids
4 Phenolic aromatic acids containing several benzene rings linked via carbon atoms (humic acids)	Humate- and humus-containing substances		Inorganic salts (such as phosphate)	Protein hydrolysates and amino acids	Plant-growth-promoting microorganisms (including mycorrhizal fungi)	Chitosan and other biopolymers	Inorganic salts
5 Vegetable oils			Seaweed extracts	Seaweed extracts		Inorganic compounds	Microorganisms
6 Natural minerals			Chitin and chitosan derivatives			Beneficial fungi	
7 Water (activated, degassed, thermal)			Antitranspirants			Beneficial bacteria	
8 Resins			Free amino acids and other N-containing substances				
9 Other raw materials (oil and petroleum fractions, shale substance)							

15 biostimulants; Ikrina and Kolbin (2004) systematized patent literature and specified 9 categories of natural raw materials used to derive biostimulants; Basak (2008) suggested that biostimulants could be grouped on the basis of single or multicomponent formulations and classified on the origin of the active ingredient, and the mode of action of the active ingredient. Du Jardin (2012) developed a scientific rationale of classification considering 8 categories of biostimulants and subsequently reduced this list to 7 categories (Du Jardin, 2015). Du Jardin (2012) was explicit in his exclusion of microorganisms from his categorization primarily to avoid conflict with existing categorization of microorganisms as biopesticides and sources of plant hormones. Later Bulgari et al. (2015) proposed a biostimulant classification on the basis of their mode of action rather than on their composition.

Many biostimulant products have been classified into completely divergent groups and categories of function, use, and type of activity (Tables 3, 4). For example, humate-based products are often described as soil health amendments while plant growth promoting rhizobacteria (PGPRs) could be categorized as biofertilizers, phytostimulators, and biopesticides (Martínez-Viveros et al., 2010; Bhattacharyya and Jha, 2012). Du Jardin (2015) has proposed that biofertilisers are a subcategory of biostimulants. Seaweed extracts have been considered as biofertilizers (Zodape, 2001) and microorganisms have also been described as biofertilizers (Vessey, 2003; Fuentes-Ramirez and Caballero-Mellado, 2006; Roy et al., 2006; Malusá et al., 2012; Bhardwaj et al., 2014; Malusá and Vassilev, 2014). Some inorganic elements or small molecules that are not known to be essential may also be classified as biostimulants if evidence of plant growth promotion is available (Michalski, 2008; Kleiber and Markiewicz, 2013; Radkowski and Radkowska, 2013). Thao and Yamakawa (2009), for example, consider phosphites to be biostimulants since plant response to phosphites frequently cannot be explained as a consequence of the known anti-fungal function of these molecules. While the categorization of biostimulants by their origin does not, *a priori*, provide information on their mode of action this categorization may still be a useful tool to aid in the process of discovery and facilitate comparison between similar products.

LEGISLATION AND LEGAL FRAMEWORK

Registration of products used in agriculture is crucial to ensure their practical, safe and legitimate application. In the absence of a sound definition of biostimulants as a discrete group of products (Basak, 2008), the registration procedure and subsequent classification regime is untenable and this inevitably creates a barrier to trade and development. Various countries, states, and administrative regions have developed different categories for registration of potential biostimulants including terminology such as plant conditioners, “other fertilizers,” supplements, soil improvers, plant strengtheners, fitofortificants, etc. (Basak, 2008; Torre et al., 2013; Traon et al., 2014). In many jurisdictions regulatory practices require an itemized description and identification of substances in all commercial product classifications while in others the registration of non-fully identified substances is allowed if those products are considered

of complex composition. There is even a proposal for complex biostimulants to not specify the chemical name (IUPAC) and note as “None” with the definition that “this product is a complex mixture of chemical substances” (Traon et al., 2014). If we accept the concept that a biostimulant is a product of clear benefit but unknown mode of action, then it can only be regulated by its safety and proof of efficacy. For example, in pharmacology it has been suggested that “the demand to demonstrate the mode of action of each single component in a phytopharmaceutical may not be obligatory any more” (Ulrich-Merzenich et al., 2009).

The complex multicomponent nature of many biostimulants clearly complicates discovery of their modes/mechanisms of action, production, registration and use. What is clearly needed however, is a regulatory mechanism to ensure that the products are “generally recognized as safe,” have “a positive benefit on crop productivity” and are discrete from existing categories of products. The task of identifying function and agronomic utility can then be pursued independently and will be driven by the marketplace imperative for product quality and consistency. Coordinating national legislation within this framework will become critical for the optimization of biostimulants and trade between different countries. The possible place of biostimulants in the regulatory system of pesticides and agrochemicals is illustrated in Figure 1.

PRIMARY SOURCES OF RAW MATERIALS

We have conducted an exhaustive analysis of the literature and categorized the majority of the reported biostimulants by origin (Table 4). Microorganisms are widely used for the production of biostimulants and may be derived from bacteria, yeasts, and fungi. These preparations may include living and/or non-living microorganisms and their metabolites. The concept of microorganism-based preparations as biostimulants is described by Xavier and Boyetchko (2002), Sofo et al. (2014), Colla et al. (2015), Matyjaszczyk (2015), and Ravensberg (2015). Different species of algae, mostly seaweeds, are also commonly used for producing biostimulants. Seaweed-based preparations as biostimulants are described in reviews by Crouch and van Staden (1993a), Khan et al. (2009), Craigie (2011), Sharma et al. (2014); and experimental papers by Goatley and Schmidt (1991), Jannin et al. (2013), Billard et al. (2014), Aremu et al. (2015b). Raw materials for biostimulants are also commonly based on higher plant parts including seeds, leaves, and roots and exudates from families *Amaryllidaceae*, *Brassicaceae*, *Ericaceae*, *Fabaceae*, *Fagaceae*, *Moringaceae*, *Plantaginaceae*, *Poaceae*, *Rosaceae*, *Solanaceae*, *Theaceae*, *Vitaceae*, among others (Naumov et al., 1993; Yakhin et al., 1998, 2011a, 2012, 2014; Pretorius, 2007, 2013; Parrado et al., 2008; Apone et al., 2010; Ertani et al., 2011a, 2013a, 2014; Colla et al., 2014; Yasmeen et al., 2014; Lucini et al., 2015; Ugolini et al., 2015). Biostimulants may also be based on protein hydrolysates and amino acids of animal origin including wastes and by-products (Mladenova et al., 1998; Maini, 2006; Kolomaznik et al., 2012; Ertani et al., 2013b; Rodríguez-Morgado et al., 2014), and insect derived chitin and chitosan derivatives (Sharp, 2013). Humate-based raw materials are widely used to derive biostimulants and have been reviewed by Sanders et al. (1990), Kelting et al. (1998), Ertani

TABLE 3 | Examples of different terminology used for commercial biostimulants.

Preparations	Source, composition	Found in the literature related terms	References
Activave®	<i>Ascophyllum nodosum</i>	Metabolic enhancer Biostimulant	Spinelli et al., 2010 Vernieri et al., 2006; Ferrante et al., 2013
Agrispon®	Natural plant extract	Biostimulant Bioregulator, Biostimulant Biostimulator	Rouse, 1984 Dubravec et al., 1995 Michalski, 2008
Aminoplant (Siapton®)	Epithelial tissues (natural substances animal origin)	Organic biostimulant, Soil fertilizer Biostimulant Fertilizer	Maini, 2006 Betti et al., 1992; Mladenova et al., 1998; Apone et al., 2006; Cambri et al., 2008; Kunicki et al., 2010 Mladenova, 1978
Asahi SL (Atonik)	Sodium para-nitrophenolate, sodium ortho-nitrophenolate, sodium 5-nitroguaiacolate	Biostimulant Bioregulator	Basak, 2008; Przybysz et al., 2014 Michalski, 2008
Bio-Algen®	<i>Phaeophyceae</i>	Biostimulator, Bioregulator	Basak, 2008
Biozyme®	<i>Ascophyllum nodosum</i> (GA ₃ +IAA+zeatin+ chelated micronutrients)	Biostimulant Bioregulator	Tandon and Dubey, 2015 Belakbir et al., 1998; Ruiz et al., 2000
ComCat®	<i>Lychnis viscaria</i>	Plant growth regulator, biostimulant,	Van der Watt and Pretorius, 2013
Ergostim®	L-cysteine and folic acid derivative	Plant growth regulator, Biostimulant Biostimulant Bioregulator, Biostimulant	Cutler and Cutler, 2004 Gupta and MacLeod, 1982; Sanders et al., 1990; Kinnersley, 1993 Dubravec et al., 1995
Fantac (Quantum)	Mixture of 5% N-Acetyl thiazolidine carboxylic acid (N-ATCA) and 0.1% folic acid	Biostimulant, growth promoter	Srivastava et al., 2008, 2010
FOLIAR (Macro-Sorb Foliar)	A complex water soluble solution derived from the enzymatic hydrolysis of animal membranes	Biofertilizer Biostimulant	Aylward, 2005 Kauffman et al., 2007
Goëmar BM 86®	<i>Ascophyllum nodosum</i>	Fertilizer, Biostimulator, Bioregulator Fertilizer	Basak, 2008 Craigie, 2011
Kelpak®	<i>Ecklonia maxima</i>	Biostimulant Biostimulator, Bioregulator Plant growth regulator, bioregulator Fertilizer Plant growth stimulant	Arthur et al., 2013; Stirk et al., 2014 Basak, 2008 Michalski, 2008 Dhargalkar and Pereira, 2005 Khan et al., 2009
Maxicrop®	<i>Ascophyllum nodosum</i>	Biostimulator, Bioregulator Fertilizer Plant growth stimulant	Basak, 2008 Dhargalkar and Pereira, 2005 Khan et al., 2009
Seasol (Agrikelp)	<i>Durvillea potatorum</i>	Plant growth stimulant Liquid organic fertiliser Biostimulant	Khan et al., 2009 Tay et al., 1987; Kurepin et al., 2014 Sharma et al., 2014
Stifun*	The complex of biologically active substances of natural origin	Bioregulator Plant growth regulator Biostimulant	Yakhin et al., 2006, 2007, 2011a Yakhin et al., 2011b, 2012, 2013 Yakhin et al., 2014, 2016a,b

(Continued)

TABLE 3 | Continued

Preparations	Source, composition	Found in the literature related terms	References
SM3 (Sea Magic 3)	<i>Laminariaceae</i> and <i>Fucaceae</i> species	Biostimulator, Bioregulator	Basak, 2008
Tytanit®	Titanium	Biostimulant Fertilizer	Basak, 2008 Kleiber and Markiewicz, 2013
Wuxal®-Ascofol	<i>Ascophyllum nodosum</i>	Biostimulator, Bioregulator	Basak, 2008
–	moringa leaf extract	Plant growth stimulant Biostimulant	Yasmeen et al., 2013 Abdalla, 2013; Yasmeen et al., 2014

*By the results of state registration tests Stifun was recommended for registration but does not registered yet.

et al. (2011b), and Jannin et al. (2012). A final category of biostimulants includes those derived from extracts of food waste or industrial waste streams, composts and compost extracts, manures, vermicompost, aquaculture residues and waste streams, and sewage treatments among others. Because of the diversity of source materials and extraction technologies, the mode of action of these products is not easily determined.

TECHNOLOGIES OF PRODUCTION

The technologies used in the production and preparation of biostimulants are highly diverse and include cultivation, extraction, fermentation, processing and purification, hydrolysis, and high-pressure cell rupture treatment (Table 4). In some instances, a biostimulant product may also contain mixes of components derived from different sources and production methods. Frequently the rationale for utilizing extracts rather than raw biomass is a consequence of the need for a standardized manufacturing process to produce a uniform commercial product (Michalak and Chojnacka, 2014). For many products, the production processes are driven by process and marketing demands and are not the result of a targeted strategy to optimize the biological efficacy of the commercial product. While the ultimate composition and possible function of commercial biostimulant products may be partially determined by both the source of raw material and the process by which it is prepared (Traon et al., 2014), there may be manufacturing processes and product treatments utilized that result in compounds that are not present in the initial (primary raw) material. An example of this is the multitude of commercial seaweed extracts, often derived from the same species, that are rarely equivalent (Craigie, 2011). Commercial biostimulant manufactured from similar sources are usually marketed as equivalent products, but may differ considerably in composition and thereby in efficiency (Lötze and Hoffman, 2016). Many manufacturers do not reveal the technology of biostimulant production since that is a commercial secret (Traon et al., 2014).

BIOACTIVE COMPONENTS AND METHODS OF QUALITY CONTROL

A diversity of substances contained in raw materials is used for the production of biostimulants. Whereas, primary metabolites are contained in most preparations *de facto*, the presence

of secondary metabolites is more specific and depends to a large extent on the raw material used (species, tissue, growing conditions). Primary metabolites include amino acids, sugars, nucleotides, and lipids (Aharoni and Galili, 2011). Secondary metabolites are formed from different primary metabolic pathways, including glycolysis, the tricarboxylic acid cycle (TCA), aliphatic amino acids (AA), the pentose-phosphate and shikimate pathways which are primarily the source of aromatic AA and phenolic compounds (PC), terpenoids/isoprenoids, nitrogen-containing compounds (alkaloids), sulfur-containing compounds (glucosinolates); (Aharoni and Galili, 2011). Frequently, biostimulants are shown to have a multicomponent composition and may include plant hormones or hormone-like substances, amino acids, betaines, peptides, proteins, sugars (carbohydrates, oligo-, and polysaccharides), aminopolysaccharides, lipids, vitamins, nucleotides or nucleosides, humic substances, beneficial elements, phenolic compounds, furostanol glycosides, sterols, etc. (Table 4). While many articles have attempted to describe the composition of complex biostimulants, these descriptions are frequently incomplete since the vast majority of biological molecules that would be present in crude extracts of complex origin, have not yet been characterized and the mere presence of a specific compound does not *a priori* demonstrate that compound is functional. The composition of most biologically derived biostimulant feed stock will also vary with the season of production, species, physiological state of the source organism and growth conditions. Indeed, there is an implication in the marketing of many biostimulants that stress conditions experienced by the plant or microbe utilized to produce the biostimulant, results in the production of stress metabolites and amino acids with consequent beneficial effects on plant response. In the absence of knowledge of the functional component of a biostimulant, changes in composition of a biostimulant over time and between batches and commercial sources cannot be interpreted. In the most rigorously prepared biostimulants from leading companies, high-throughput analytical methods have been employed to ensure consistent product quality (Sharma et al., 2012b). Methods such as chromatography, mass spectrometry, NMR spectroscopy, elemental analysis, ELISA, spectrophotometry, etc. are typically used for this purpose (Table 4). The complexity of this challenge is illustrated in the analysis of a four-year algae composition sequence using a profile or fingerprint technique employing NMR (Craigie et al., 2009).

TABLE 4 | Biostimulants: sources, production, compositions, and activities.

Genus, species of organism /source of raw material	Methods of production	Methods of identification/standardization	Ingredients and bioactive compounds	Hypothesized modes/mechanism of action	Biological effects
1	2	3	4	5	6
1. BACTERIA					
1. Preparations of living microorganisms:					
<i>Aeromonas rivuli</i> , <i>Agromyces fucosus</i> , <i>Bacillus licheniformis</i> , <i>Bacillus megaterium</i> , <i>Bacillus pumilus</i> , <i>Bacillus safensis</i> , <i>Microbacterium sp.</i> , <i>Nocardia globberula</i> , <i>Pseudomonas fluorescens</i> , <i>Pseudomonas tulva</i> , <i>Pseudoxanthomonas dajemonensis</i> , <i>Rhodococcus corynophilus</i> , <i>Sphingopyxis macrogoltabida</i> , <i>Streptomyces sp.</i>	Cultivation	ARISA fingerprinting, ELISA, GC-MS, Immunoblot, Most Probable Number, NMR, Spectroscopy, Molecular taxonomical characterization; Plate Count methods, Thermochemolysis, TLC.	Substances with auxin (IAA)-like bioactivity, IAA, cytokinins, betaines, gibberellins, amino acids, oligopeptides, low-molecular-weight peptides, peptidoglycans; lypopolysaccharides, melatonin.	Increase availability of nutrients in soil. Stimulate nitrogen uptake. Maintain soil fertility, nitrogen fixation, solubilize insoluble minerals through the production of organic acids. Auxin-like, gibberellin-like activity. Influence on the hormonal status of the plant. Stimulate amino acid synthesis. Increase concentrations of total carbohydrates. Increase nutrients (magnesium, nitrogen and phosphorus, etc.). Increase pigments (chlorophyll, carotenoids). Increase antioxidant substances. Stress resistance: heat, drought, wear, traffic, and/or salinity. Control fungal diseases and other physiological disorders. Activation of systemic resistance (ISR and SAR).	Increase germination rate, growth characters (length, fresh, dry weight) of shoots and roots, plant quality, productivity, yield.
2. Preparations on the basis of non-living microorganisms and their metabolites:					
<i>Bifidobacterium bifidus</i> , <i>Lactobacillus sp.</i> , <i>Lactobacillus acidophilus</i> , <i>Lactobacillus buchneri</i> , <i>Lactobacillus delbrueckii</i> , <i>Lactobacillus johnsonii</i> , <i>Lactobacillus murinus</i> , <i>Lactobacillus paraplantarum</i> , <i>Lactobacillus pentosus</i> , <i>Lactobacillus plantarum</i> , <i>Lactococcus lactis</i> , <i>Leuconostoc oenos</i> , <i>Propionibacterium freudenreichii</i> , <i>Propionibacterium pelophilus</i> , <i>Propionibacterium shermanii</i> , <i>Propionibacterium spp.</i> , <i>Propionivibrio limicola</i> , <i>Streptococcus spp.</i> , <i>Streptococcus thermophilus</i> (also called <i>Streptococcus salivarius</i>); Bacterial cell cream from an industrial fermentation process.	Acid hydrolysis, alkali hydrolysis, cultivation, enzymatic hydrolysis, fermentation.				
References: Bashan, 1998; Inseer et al., 2006; Borriss, 2011; Tachibana et al., 2012; Abbas, 2013; de Fretes et al., 2013; Giannattasio et al., 2013; Janas and Posmyk, 2013; Jenkins, 2014; Sofo et al., 2014; Colla et al., 2015; Spaepen, 2015.					
2. FUNGI					
1. Preparations of living microorganisms:					
<i>Glomus intraradices</i> , <i>Trichoderma atroviride</i> .	Cultivation, fermentation, lyophilization.	ARISA fingerprinting, sonication and gradient filtration, NMR, bioassays, HPLC, FTIR.	Amino acids, auxin-like compounds, betaines, carbohydrates, chitosan, cytokinins, exopolysaccharides, gibberellins, IAA, melatonin, minerals, nucleic acids, oligopeptides, oligoproteins, polyglucuronic acid, proteins, siderophores, vitamins.	Increase nutrient uptake. Stimulate of nitrogen uptake. Increase enzyme activity. Influence on soil and plant metabolism. Change hormonal status of the plant. Stimulate amino acid synthesis. Increase total carbohydrates and total protein. Increase in total soluble sugars, total free amino acids, and total phenols. Increase pigments (chlorophyll, carotenoids). Increase nutrients concentrations. To induce plant defense reactions. Enhance environmental stress tolerance: drought, salinity, soil disturbance, toxic pollutants. Limit spread of disease by microbial competition. Prevent pathogen infection by eliciting resistance mechanisms such as systemic induced resistance. Reduce pathogen inoculum in the rhizosphere, thereby reduce the incidence of infection.	Increase germination rate, growth characters (length, fresh, dry weight) of shoots and roots; vegetative growth; the size of plants; the number of flowers; the number of fruits; plant quality; productivity; yield and yield components.
2. Preparations derived from non-living microorganisms and their metabolites:					
<i>Candida spp.</i> , <i>Hanseniaspora spp.</i> , <i>Issatchenkia spp.</i> , <i>Kloeckera spp.</i> , <i>Kluyveromyces spp.</i> , <i>Meischnikowia spp.</i> , <i>Pichia spp.</i> , <i>Saccharomyces bayanus</i> , <i>Saccharomyces boulardii</i> , <i>Saccharomyces cerevisiae</i> , <i>Saccharomyces exiguus</i> , <i>Saccharomyces pastorianus</i> , <i>Saccharomyces pombe</i> , <i>Syncephalastrum racemosum</i> .					
References: Xavier and Boyetchiko, 2002; Acholeya et al., 2005; Udo et al., 2010; Gandarillas Infante, 2012; Abbas, 2013; Giannattasio et al., 2013; Janas and Posmyk, 2013; Hammad and Ali, 2014; Jenkins, 2014; Sofo et al., 2014; Valepyn et al., 2014; Colla et al., 2015.					

(Continued)

TABLE 4 | Continued

Genus, species of organism /source of raw material	2	3	4	5	6
1					
3. ALGAE					
<i>Ascophyllum nodosum</i> , <i>Caulerpa scapelliformis</i> , <i>Chlorella ellipsoida</i> , <i>Dunillaea antarctica</i> , <i>Dunillaea potatorum</i> , <i>Ecklonia maxima</i> , <i>Enteromorpha flexuosa</i> , <i>Fucus serratus</i> , <i>Fucus vesiculosus</i> , <i>Gelidium acerosa</i> , <i>Gracilaria corticata</i> , <i>Gracilaria salicornia</i> , <i>Himantalia elongata</i> , <i>Hypnea musciformis</i> , <i>Kappaphycus alvarezii</i> , <i>Laminaria digitata</i> , <i>Laminaria hyperborea</i> , <i>Macrocystis integrifolia</i> , <i>Macrocystis pyrifera</i> , <i>Padina boergeseni</i> , <i>Padina gymnospora</i> , <i>Padina pavonica</i> , <i>Sargassum muticum</i> , <i>Sargassum tenerum</i> , <i>Sargassum wightii</i> , <i>Spirulina maxima</i> , <i>Ulva lactuca</i> .	Acid processing; acidic extraction; alkaline extraction; alkaline hydrolysis; alkaline processing; aqueous extraction; cell burst; cell rupture with high pressure treatment; cold or frozen, alkaline and water extractions; cryoprocessing; enzyme-assisted extraction (EAE); fermentation; heated alkaline hydrolysis; microwave-assisted extraction (MAE); neutral extraction; pressurized liquid extraction (PLE); also known as pressurized fluid extraction, enhanced solvent extraction, high-pressure solvent extraction, or accelerated solvent extraction techniques; supercritical fluid extraction (SFE); ultrasound-assisted extraction (UAE).	¹³ C NMR, ¹ H NMR (qNMR), Bioassay, DEPT together with 2D experiments (COSY, GHSQC and GHMBC), ELISA, energy dispersive X-ray microanalysis (EDX), Fourier-transform infrared spectroscopy (FTIR), GC/MS, GLC, HPLC, HPLC/MS/MS, HPLC/MS, inductively coupled plasma-optical emission spectroscopy (ICP-OES), IR, LC-MS, LC-MS-MS, mass spectrometry (ESI-TOF-MS), NMR, pyrolysis gas chromatography/mass spectrometry (Py-GC/MS), scanning electron microscopy, thermogravimetry (TGA), TLC, Ultra high performance liquid chromatography–tandem mass spectrometry (UHPLC-MS/MS) analysis, X-ray microanalysis.	1-Aminocyclopropane-1-carboxylic acid (ACC); abscisic acid (ABA); alginic acid; Auxins (IAA, IAAsp, IAAa, IAGly, IALeu, ICA, ILA, IPA, IPIa, ICA, N,N-dimethyltryptamine, IALD, iso-indole, 1, 3-dione (N-hydroxy ethylphthalimide), auxin-like substances, phenyl-3-acetic acid (PAA) and hydroxyphenyl acetic acid (OH-PAA); Betaines (Glycinebetaine, γ-aminobutyric acid betaine, δ-aminovaleic acid betaine, glycinebetaine, laminine, lysinebetaine, ascorbylline); Carbohydrates: 1-(2-furanyl) ethanolone (mannitol), 5-methyl-2-furcarboxaldehyde (fucoidan), 2-hydroxy-3-methyl-2-cyclopenten-1-one (laminarin), dihydrodromannitol (mannitol), 1,6-anhydromannopyranose and 1,6-anhydromannofuranose (mannitol); Cytokins: zeatin (Z), dihydrozeatin (DHZ), trans-zeatin (tZR), cis-zeatin (cZR), dihydrozeatin riboside (DHZR), isopentenyladenine (IP), isopentenyladenosine (IPR), benzyladenine riboside (BAR), meta-topolin (mT), meta-topolin riboside (mTR), ortho topolin (oT), and ortho-topolin riboside (oTR), cytokinin glucosides, etc. Gibberellin acid (GA ₃); carrageenans; lipids; melatonin; minerals (Na, Cu, Fe, I, K, Mg, Na, P, S, B, Mn, Zn, Co, potassium oxide, phosphorus oxide, N, S, C, HCO ₃ ⁻ , etc.); oligosaccharides; pepsin; phenolic compounds; eckol, phloroglucinol, etc. polysaccharides, protein, sterols; 22-Dehydrocholesterol; 24-Methylecholesterol; 24-Methylenecycloartanol; 24-Methylenophenol; 28-Isotucosterol; 5-Dihydroergosterol; Brassicasterol;	Increase nutrient absorption and fertilizer efficiency; nutrient uptake; uptake of Cu, Ca, K and Mg; macro- and microelements content; assimilation of N, C, and S; could reduce the fertilizers. Efficient water uptake. Auxin- Cytokinin- Gibberellin-like activity. Modulation of phytohormones. Regulation of gene expression. Increase photosynthetic efficiency; photosynthetic pigments (chlorophyll, carotenoids); total protein concentrations; amino acid, betaines, carbohydrate content; ascorbic acid; nutrient concentrations. Increase metabolites including phenolic compounds. Up-regulation of bio-synthetic enzymes; enhance antioxidant activity. Enhance biosynthesis of non-enzymatic compounds. Delay senescence. Reduce transpiration; Enhance stomatal conductance; Change of metabolism; Alter of root architecture; Modulation of root exudates; Activate the mechanisms of strengthening cell walls. Decrease rate of transpiration; sensitivity of the plants to water deficiency. Resistance to frost, insect and pathogen attack, disease and pests; enhance locally plant immunity against viruses; reduced virus infection; reduction in root-knot nematode infestation; against salinity stress; water stress; induce improvement of plant growth under sea water stress. Tolerant to iron deficiency.	Increase number of fruits per plant and size of fruit; fruit and crop yield; fruit quality; development of a vigorous root system and improved growth; increase in fresh weight, grain weight and yield components; root formation; growth characters (length, fresh, dry weight) of shoots and roots; quality of the plants; stimulate the growth; induce rooting.

(Continued)

TABLE 4 | Continued

Genus, species of organism /source of raw material	Methods of production	Methods of identification/standardization	Ingredients and bioactive compounds	Hypothesized modes/mechanism of action	Biological effects
1	2	3	4	5	6
			Campesterol; Cholesterol; Chondrillasterol; Clerosterol; Cionasterol; Codisterol; Cycloartenol; Decortinol; Decorinone; Desmosterol; Ergosterol; Fucosterol; Isodecortinol; Obusifolol; Ostreasterol; Poriferasterol; Sitosterol; β -Sitosterol; Stigmasterol; Zymosterol; $\Delta^{4,5}$ — Ketosteroids; Δ^5 — Ergosterol; Δ^7 — Ergosterol, etc.		
<p>References: Aitken and Senn, 1985; Featonby-Smith and van Staden, 1983; Finnie and van Staden, 1983; Painter, 1983; Nelson and van Staden, 1985; Sanderson and Jameson, 1986; Hirsch et al., 1989; Crouch et al., 1992; Verkleij, 1992; Crouch and van Staden, 1993a,b; Nabil and Cosson, 1996; Stirk and Van Staden, 1996; Stirk et al., 2001; Stirk et al., 2003, 2014; Vernieri et al., 2005; Abd El-Baky et al., 2008; Leal et al., 2008; Rayorath et al., 2008; Khan et al., 2009, 2011; Vera et al., 2011; Zodape et al., 2012a,b, 2014; Abbas, 2013; Arthur et al., 2013; Chbani et al., 2013; Ghamam et al., 2013; González et al., 2013; Janas and Posmyk, 2013; Jannin et al., 2013; Thomas et al., 2013; Brice-o-Dominguez et al., 2014; Hernandez-Herrera et al., 2014; Michalak and Chojnacka, 2014; Mikiciuk and Dobromilska, 2014; Petrozza et al., 2014; Stadnik and de Freitas, 2014; Vijayanand et al., 2014; Vrioth et al., 2014; Arenu et al., 2015b; Omar et al., 2015; Rengasamy et al., 2015a,b; Satish et al., 2015.</p>					
4. HIGHER PLANTS					
<i>Agapanthus africanus</i> , <i>Allium sativum</i> , <i>Brassica juncea</i> , <i>Brassicaceae</i> , <i>Camellia sinensis</i> , <i>Castanea sativa</i> , <i>Ceratania silqua</i> , <i>Digitalis sp.</i> , <i>Fabaceae</i> , <i>Helianthus annuus</i> , <i>Lupinus albus</i> , <i>Lupinus sp.</i> , <i>Lycopersicon esculentum</i> , <i>Lycopersicon sp.</i> , <i>Malus sp.</i> , <i>Medicago sativa</i> , <i>Moringa oleifera</i> , <i>Musa acuminata</i> , <i>Nicotiana tabacum</i> , <i>Quercus sessiliflora</i> , <i>Saccharum officinarum</i> , <i>Vaccinium sp.</i> , <i>Vitis vinifera</i> , <i>Zea mays</i> .	Alkaline hydrolysis; aqueous extraction; controlled fermentation; conventional solid-liquid extraction; cool extraction; fully controlled enzymatic hydrolysis; ethanol extraction; microwave extraction; pressurized solvent extraction; solid-liquid dynamic extraction.	^{13}C NMR; ^1H NMR; Bioassays; C, N elemental analysis; column chromatography fractionation; COSY; DEPT; ELISA; FT-IR; GC-MS; HPLC; HMBC; HMQC; HPLC-DAD-MS; HS-SBSE-GC-MS; ICP-OES; liquid-solid extraction; mass spectrometry; NOESY; preparative thin layer chromatography; qualitative thin layer chromatography; Raman spectroscopy; spectra analysis.	Amino acids: alanine; arginine; aspartic acid; cysteine; glutamic acid; glycine; histidine; isoleucine; leucine; lysine; methionine; phenylalanine; proline; serine; threonine; tryptophan; tyrosine; valine, etc. Auxins: indole-3-acetic acid (IAA), indolbutyric acid (IBA), Naphthoxy acetic acid (NAA); carbohydrates: galactose, glucose, mannose, xylose, arabinose, cellulose, catalase; cytokinins: isopentenyladenosine (IPA), kinetin, etc. elements (N, P, K, Na, S, K, Ca, Mg, P, B, Fe, Zn, Cu, Mn, Ni, Cl, Mo, Co, etc.) ellagitannins: Castalagin; Vescalagin; Roburin E; Grandinin + roburin D; Roburin A + B; Roburin C Enzymatic antioxidants: flavanols (catechin, epicatechin), flavonoid compound: 3-[[O- β -D-glucopyranosyl-(1'-'-3')- α -L-rhamnosyl-(1'-'-2')- β -D-glucopyranosyl]oxy] agapantheinin, flavanone, naringenin (5,7,4'-trihydroxyflavanone),	Increase of nitrogen assimilation. Increase phosphate uptake. Induction of morphological changes in root architecture. Decrease in accumulation of Na ⁺ and Cl ⁻ into shoots under moderate saline conditions. Auxin-, cytokinin-, gibberellin-like activity. Regulation of hormonal system. Regulation of gene expression. Improve photosynthetic rate. Increasing the efficiency of light utilization and dissipation of excitation energy in the PSII antennae. Increase chlorophyll and carotenoids content. Stimulate plant nitrogen and carbon metabolisms. Increase of functional activity of nucleoli of meristematic cells. Increase biochemical contents: leaf nitrogen content; protein amount; free amino acids; carbohydrates; the total sugars; contents of lectin; NADP ⁺ ; an increase in phenolics in plant tissues; ascorbic acid, β -carotene, elements (N, P, K, Ca, Mg, Fe, etc.). Higher concentrations of macronutrients in the plant tissue. Increase in osmolytes. Changes in sterols, terpenes, glucosinolates composition. Increase SPAD index. Regulation of enzyme activity. Modulating enzymatic and non-enzymatic antioxidant systems. Effects on phenylpropanoid metabolism. Activation of antioxidant defense system. Improved water use efficiency. Regulation of stomata. Enhance plant resistance to stress conditions. Significant antimicrobial activity, especially antifungal activity.	Growth stimulating effect. Promoting plant growth, health or yield. Increase seed germination; in coleoptile elongation rate; plant biomass, the shoot, root dry weight, root length, and root area, the total dry biomass, plant growth. Ability to manipulate early seedling growth. Rooting. Plant height, number of flowers and number of fruits per plant. Maturity. Enhance the yield of vegetable crops. Strong positive effects on growth, development and fruit quality. Increase in the organoleptic and quality food parameters.

(Continued)

TABLE 4 | Continued

Genus, species of organism /source of raw material	Methods of production	Methods of identification/standardization	Ingredients and bioactive compounds	Hypothesized modes/mechanism of action	Biological effects
1	2	3	4	5	6
			5, 7, 3', 4'-tetra-O-acetylflavanone, trans-4, 2', 4'-tri-O-acetylchalcone – Isoliquiritigenin folic acid, free enzymatic proteins furostanol glycosides gibberellins; gibberellic acid; Gibberellin A4 + A7; glycosides, humic acids inositol lignin and hemicellulose moieties, Lipids, Low molecular weight polyphenols: vanillin; syringaldehyde; coniferaldehyde; sinapaldehyde; vanillic acid; syringic acid; gallic acid; ellagic acid, Melatonin nucleosides: purine, pyrimidine, nucleotides, oligosaccharides organic acids organic nitrogen and organic carbon peptides, Peroxidase, phenolic acids: trans-caftaric acid; trans-p-coumaric acid; ferulic acid; trans-caftaric acid; trans-p-coumaric acid; ellagic acid; gallic acid; protocatechuic acid, syringic acid, vanillic acid), stilbenes (piceid, trans-resveratrol), pyrogallol, sinapaldehyde, Polyphenols polysaccharides, protein, saponins; sugars, superoxide dismutase; tannins triacontanol (TRIA), triglyceride vitamins (A, B1, B2, B3, B6 and PP, C, E) Volatile composition: acids (hexanoic, octanoic, decanoic), alcohols (1-hexanol, 3-hexen-1-ol, 1-octen-3-ol, 1-nonanol, linalool, α -terpineol, guaiacol, benzyl alcohol, 2-phenylethanol), aldehydes (benzaldehyde, nonanal, vanillin), furanic compounds (furfural, 5-hydroxymethylfurfural, 5-methylfurfural, 2-furanmethanol, methyl furoate), lactones (trans-whiskey lactone, cis-whiskey lactone), D-limonene, geranyl acetone, linalyl acetate, β -ionone, stilbene 1,2. Volatile compounds:	Reduce numbers of root knot nematodes. Enhance plant resistance to nematodes. Alleviate the effect of drought; salinity; ameliorate salinity-induced adverse effects.	

(Continued)

TABLE 4 | Continued

Genus, species of organism /source of raw material	Methods of production	Methods of identification/standardization	Ingredients and bioactive compounds	Hypothesized modes/mechanism of action	Biological effects
1	2	3	4	5	6
			5-hydroxymethylfurfural; 6-methoxyeugenol; acetovanillone; benzaldehyde; cis- β -methyl- γ -octalactone; eugenol; furalal; guaiacol; trans- β -methyl- γ -octalactone.		
References: Murch and Saxena, 2002; Ciesiolka et al., 2005; Fleming et al., 2006; Parrado et al., 2007, 2008; Pretorius, 2007, 2013; Virji, 2007; Schiavon et al., 2008; Erfani et al., 2009, 2011b, 2013b, 2014; Vyas et al., 2009; Apone et al., 2010; Rivera et al., 2010; Paradiković et al., 2011; Van der Watt and Pretorius, 2011; Yakhin et al., 2011a,b, 2012; De Lucia and Vecchiotti, 2012; Hanafy et al., 2012; Abdalla, 2013; Bargiacchi et al., 2013; Christofolotti et al., 2013; Daniels, 2013; Janas and Posmyk, 2013; Pretorius, 2013; Yasmine et al., 2013, 2014; Ziosi et al., 2013; Arnao and Hernández-Ruiz, 2014; Baglieri et al., 2014; Caulier et al., 2014; Chambers, 2014; Colla et al., 2014; Lachhab et al., 2014; Sánchez-Gómez et al., 2014; Lucini et al., 2015; Pardo-García et al., 2014; Ugolini et al., 2015.					
5. ANIMAL RAW MATERIALS					
Animal epithelium, by-products deriving from leather manufacture, chicken feathers, chitin-containing waste materials from the seafood industry, epithelial tissue, hemoglobin hydrolysate, hydrolysis of chrome-tanned waste, leather waste by enzymatic hydrolysis, meat flour, secondary processing of leather waste materials - complex process of collagen protein hydrolysis gained from tannery wastes, waste bovine hooves and horns.	Acid hydrolysis; chemical hydrolytic processes; controlled hydrolysis; Enzymatic hydrolysis; thermal hydrolytic processes.	Amino acid analysis; Bioassays; ecotoxicological tests; Fourier transform infrared spectroscopy; gas chromatography coupled with mass spectrometry(GC/MS); sodium dodecyl sulfate polyacrylamide gel electrophoresis.	elements (Na, S, K, Ca, Mg, P, Fe, Zn, Cu, Mn, Ni, B, etc.); fat; free amino acids (aspartic acid, hydroxyproline, threonine, serine, glutamic acid, proline, glycine, alanine, valine, methionine, isoleucine, leucine, tyrosine, phenylalanine, γ -aminobutyric acid, histidine, ornithine, lysine, arginine, cysteine, cystine, etc.); melatonin; organic matter; peptides; protein; short-chain peptide bound amino acids.	Improve the utilization of nutrients in plants. Induce morphological changes in root architecture. Auxin-, cytokinin-, gibberellin-like activity. Change hormone levels. Affect on biochemical systems that regulate the biosynthesis of natural plant growth regulators. Synergistic effect with exogenous PGR. Induction of gene expression. Increase of enzyme activities. Accelerate major metabolic reactions. Photosynthetic rate. Increase pigment content, proteins, vitamin C, phenolic contents. Enhance contents of potassium, sodium, copper, zinc and iron in vegetal tissues; alter stomatal conductance; CO ₂ assimilation; reduce transpiration. Increase enzymatic activities and soil biodiversity. Anti-stress effect under drought, high temperatures and freezing, mechanical and chemical stress, viral infection. Stimulate the growth and activity of beneficial microbes. Improve antioxidant activity.	Better root growth and development, effects on foliar growth. Increase root and leaf growth. Root formation. Induction of flowering. Improve good fruit setting and reduce fruit drop. Make more uniform fruit weight and size. High yields.
References: Miller et al., 1955; Weissbach et al., 1959; Mladenova, 1978; Betti et al., 1992; Mladenova et al., 1998; Murch and Saxena, 2002; Khan et al., 2003; Apone et al., 2006; Maini, 2006; Kauffman et al., 2007; Cambri et al., 2008; Parrado et al., 2008; Erfani et al., 2009, 2013a; Veselá and Friedrich, 2009; Lisiecka et al., 2011; Paradiković et al., 2011; De Lucia and Vecchiotti, 2012; Kolomaznik et al., 2012; Pecha et al., 2012; Janas and Posmyk, 2013; Migliore et al., 2013; Paradiković et al., 2013; Vaskova et al., 2013; Arnao and Hernández-Ruiz, 2014; Corte et al., 2014; Rodríguez-Morgado et al., 2014; Lachhab et al., 2014.					
6. HUMATE-CONTAINING RAW MATERIALS					
Compost, humic-like substances extracted from agro-industrial wastes, Leonardite, lignin, peat, soil, vermicompost, volcanic soil, waste materials.	Extraction; thermochemolysis.	¹³ C NMR; ¹ H NMR; atmospheric pressure chemical ionization-mass spectrometry (APCI-MS); bioassays; CP/MAS; cross-polarization magic angle spinning (CPMAS) - ¹³ C-NMR; diffuse-reflectance infrared Fourier transform spectroscopy (DRIFT); electronic microscopy; elemental analysis; FTIR; HPLC/MS/MS; HPSEC; pyrolysis-gas chromatography-mass spectrometry; UV-vis.	amino acids; cellulose and hemicelluloses and saccharides; elements: C, H, N, O; Ca, Cu, Fe, K, Na, P, S, Si, Zn, etc. fatty acids; flavonoids; high-molecular humic substances; humate potassium acid/fulvic acid; humic substances; lignins; lipids; microorganisms; peptides; phenolic acids (protocatechuic acid, p-hydroxybenzoic acid, p-coumaric acid, ferulic acid; phenols; plant hormones:	Induce NO (Nitrous Oxide) synthesis. Increase nitrate uptake. Enhance nutrient uptake and nutritional status. Increase root-to-shoot translocation of elements. Auxin-, cytokinin-, gibberellin-like activity. Regulate of hormonal status. Regulate of gene expression. Ability stimulate various metabolic pathways. Changes in primary and secondary metabolism. Increased chlorophyll a, b and total carotene content. Regulation of photosynthesis, carbon (C) metabolism. Increase assimilation of N, C, and S. Increase protein; phenol content, polyamines. Stimulate the activity of enzymes. Enhance phenylpropanoid metabolism.	The activation of growing processes. Increase root and leaf growth. Increase growth characters (length, fresh, dry weight) of shoots and roots. Increase root size, branching. General increase of biomass. Increase total and marketable yields.

(Continued)

TABLE 4 | Continued

Genus, species of organism /source of raw material	Methods of production	Methods of identification/standardization	Ingredients and bioactive compounds	Hypothesized modes/mechanism of action	Biological effects
1	2	3	4	5	6
			auxin (IAA); Brassinosteroids (Brassinolide, Castasterone, Teasterone, Typhasterol, 28-Homocasterone, Cathasterone); Cytokinins (Z, tZR, tZRMF, tZOG, tZROG, cZ, cZR, cZRMF, cZOG, cZROG, DHZRMF, DHZROG, IP, IPR, IPRMP); gibberellins (GA ₁ , GA ₂ , GA ₃ , GA ₄ , GA ₅ , GA ₆ , GA ₇ , GA ₈ , GA ₉ , GA ₁₃ , GA ₁₅ , GA ₁₉ , GA ₂₀ , GA ₂₄ , GA ₂₉ , GA ₃₄ , GA ₄₄ , GA ₆₁ , GA ₆₃); proteins.	Alter REDOX homeostasis. Enhance water, salinity and heavy metal stress tolerance. Changes on root architecture. Stimulate of chloroplast division. Alter microorganism communities in the rhizosphere.	

Reference: Cacco and Dell'Agnola, 1984; Sanders et al., 1990; Russo and Berlyn, 1991; Adani et al., 1998; Zhang and Schmidt, 2000; Canellas et al., 2002, 2010; Chen et al., 2004; Chambolle, 2005; Nardi et al., 2005, 2006, 2007; Zandonadi et al., 2009; Vasconcelos et al., 2009; Dobbss et al., 2010; Mora et al., 2010; Schiavon et al., 2010; Trevisan et al., 2010; Ertani et al., 2011a, 2013c; Aydin et al., 2012; Garcia et al., 2012; Jannin et al., 2014; Abbas, 2013; Pizzeghello et al., 2013; Berbara and Garcia, 2014; Billard et al., 2014; Canellas and Olivares, 2014; Arenu et al., 2015a; Hernandez et al., 2015.

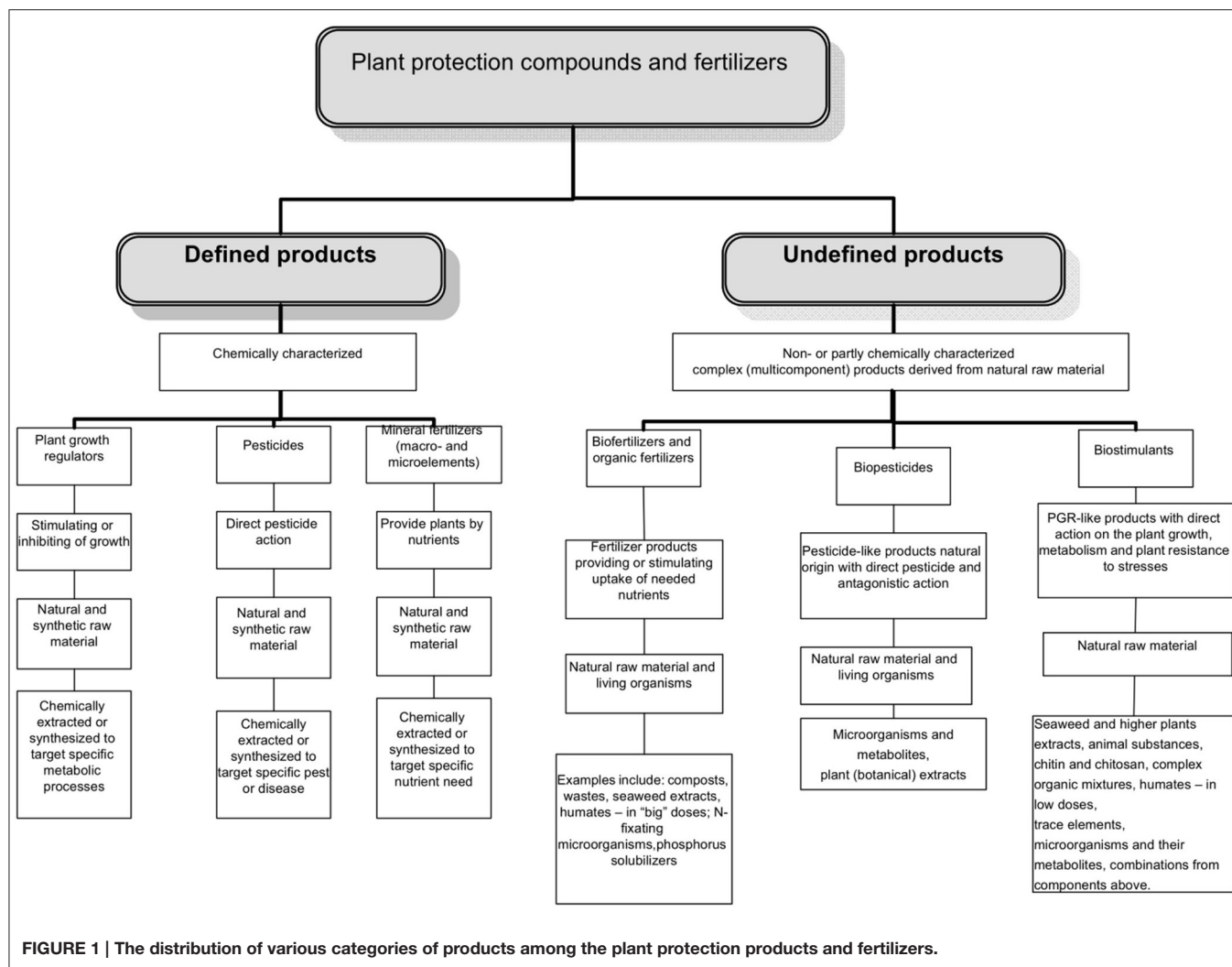
FUNCTION AND EFFECTS ON WHOLE PLANTS

Biostimulants have been used at all stages of agricultural production including as seed treatments, as foliar sprays during growth and on harvested products. The mode/mechanisms action of “biostimulants” is equally diverse and may include the activation of nitrogen metabolism or phosphorus release from soils, generic stimulation of soil microbial activity or stimulation of root growth and enhanced plant establishment. Various biostimulants have been reported to stimulate plant growth by increasing plant metabolism, stimulating germination, enhancing photosynthesis, and increasing the absorption of nutrients from the soil thereby increasing plant productivity (Table 4). Biostimulants may also mitigate the negative effects of abiotic stress factors on plants and marked effects of biostimulants on the control of drought, heat, salinity, chilling, frost, oxidative, mechanical, and chemical stress, have been observed (Table 4). Alleviation of abiotic stress is perhaps the most frequently cited benefit of biostimulant formulations. The following text describes the primary modes/mechanisms of action that have been demonstrated or claimed for biostimulants in the primary scientific literature.

MODES OF ACTION/MECHANISMS OF ACTION

Understanding the modes of action of an agricultural chemical has been a fundamental requirement for effective marketing and frequently a regulatory requirement for manufactured products used in agriculture. Mode of action is used here to mean “a specific effect on a discrete biochemical or regulatory process,” thus the “mode of action” of Glyphosate is to inhibit the activity of the enzyme enolpyruvylshikimate-3-phosphate synthase (EPSPS). Biostimulants frequently do not meet this standard of specificity and indeed there are few biostimulant products for which a specific biochemical target site and known mode of action has been identified. For a small subset of biostimulants, however, a demonstrated impact on general biochemical or molecular pathways or physiological processes, termed here as a “mechanism of action,” has been identified even though the explicit “mode of action” may not be known. An example of a “mechanism of action” would be a stimulation of photosynthesis or the down regulation of a plant stress signaling pathway without an understanding of the explicit biochemical or molecular “mode of action.”

For many biostimulant products, however, neither a specified mode of action, nor a known mechanism of action, has been identified. The presence of some spurious products in the marketplace compromises the market for all players resulting in the assumption by many, that biostimulants as a whole, are “snake oils” (Basak, 2008), a pejorative term implying the product is of no value. Multicomponent biostimulants are particularly difficult to reconcile since they may have constituents for which the mode of action is known and components of no known functional benefit. Furthermore, multicomponent biostimulants



will frequently contain measureable but biologically irrelevant concentrations of known essential elements, amino acids, and plant hormones etc., for which the mode of action is known but the concentrations are irrelevant when used at recommended rates. Thus, for many of the multicomponent biostimulant in the marketplace today, we propose that a demonstration of a clear “mechanism of action” is a more rationale and attainable regulatory goal than requiring an unequivocal demonstration of the “mode of action.”

Insight into the use of the terms “mode and mechanism” of action can be drawn from the pesticide science and pesticide development. In pesticide science, the “mechanism of action” describes the integral of all the biochemical events following application while the “mode of action” characterizes the main features of a bioactive molecule and its specific biochemical action leading to its effect in treated plants (Aliferis and Jabaji, 2011). In reference to plant bioregulators, Halmann (1990) suggests that ideally an understanding of the mode of action of plant bioregulators on the molecular level requires the identification of the receptor site for each regulator,

as well as the elucidation of the subsequent reactions. In reality this standard is often not met in biopesticide or biostimulant products where the identification of the molecular targets of all bioavailable (and frequently uncharacterized) compounds within a given extract cannot be easily achieved. The identification of the target binding sites of the natural biomolecules has, however, proven to be helpful in the design of new insecticidal molecules with novel modes of action (Rattan, 2010).

At the present time, given the difficulty in determining a “mode of action” for a complex multicomponent product such as a biostimulant, and recognizing the need for the market in biostimulants to attain legitimacy, we suggest that the focus of biostimulant research and validation should be upon determining the mechanism of action, without a requirement for the determination of a mode of action. This is the standard of practice for many pharmacological products. With the development of advanced analytical equipment, bioinformatics, systems biology and other fundamentally new methodologies a more complete understanding of the mechanisms and even

possible modes of action of these materials may be achieved in the future. While this proposal suggests that the development and marketing of a biostimulant may not require a demonstration of the mode of action, it is still in the interest of the manufacturers of these products to pursue an understanding of the mode of action so that the product can be improved and the use can be optimized for various environments and cropping systems.

The mechanisms of action of all but a few biostimulants remain largely unknown (Rayorath et al., 2008; Khan et al., 2009; Rathore et al., 2009). This is primarily due to the heterogeneous nature of raw materials used for production and the complex mixtures of components contained in biostimulant products which makes it almost impossible to identify exactly the component(s) responsible for biological activity and to determine the involved mode(s) of action (Paradićević et al., 2011). Therefore, focus should be upon the identification of the “mechanisms of action” of biostimulants as indicated by general positive impacts on plant productivity through enhancement in processes such as photosynthesis, senescence, modulation of phytohormones, uptake of nutrients and water, and activation of genes responsible for resistance to abiotic stresses and altered plant architecture and phenology (Khan et al., 2009; Sharma et al., 2012b). An example of this process is the advances in use of protein-based biostimulants for which recent studies have identified the target metabolic pathways and some of the mechanisms through which they exert their effects on plants (Nardi et al., 2016).

To further our understanding of modes/mechanisms of biostimulant action we have systematized the stages of biostimulants action on plants after their application: (1) penetration into tissues, translocation and transformation in plants, (2) gene expression, plant signaling and the regulation of hormonal status, (3) metabolic processes and integrated whole plant effects.

PENETRATION INTO TISSUES, TRANSLOCATION, TRANSFORMATION IN PLANTS

The penetration of amino acids and peptide based biostimulants into plant tissues has been investigated using radiolabeled amino acids (Maini, 2006) and mathematical modeling (Kolomazník et al., 2012; Pecha et al., 2012). The components of a biostimulant preparation of animal origin, labeled with ^{14}C proline and glycine, were shown to penetrate rapidly into treated leaves and where subsequently distributed to other leaves (Maini, 2006). The mathematical model based on the “mechanism of diffusion” allows the estimation of the time required for the absorption of a minimal amount of the active component of a biostimulant. Furthermore, it describes the process of its transport from the moment of penetration into the leaf until the arrival at more distant tissues (Kolomazník et al., 2012; Pecha et al., 2012). The penetration of protein hydrolysates into a plant tissue occurs via diffusion of protein molecules through membrane pores (Kolomazník et al., 2012) and is energy-dependent (Parrado et al.,

2008). Biostimulants must have a good solubility in water or other suitable solvents. This is a precondition for most types of application and for sufficient penetration of active ingredients into internal structures of treated plants. Surfactants and other additives may be required to overcome solubility and uptake limitations including lipophilicity and molecular size of active components (Kolomazník et al., 2012; Pecha et al., 2012).

GENE EXPRESSION, SIGNALING, AND HORMONE INTERACTIONS

Ultimately a full understanding of the biological activity of complex biostimulant preparations will require a detailed understanding of the mechanism of action and effects on plant productivity and the identification of the biologically active molecules and their molecular mode of action (Henda and Bordenave-Juchereau, 2014). A wide array of molecular methods has been used to attempt to discern the active compounds found in biostimulants including microarrays, metabolomics, proteomic, and transcriptomics methods. These technologies have been applied to biostimulants to probe changes in gene expression following the application of biostimulants (Jannin et al., 2012, 2013; Santaniello et al., 2013). Further research on the effects of complex biostimulants and their components on the complete genome/transcriptome of plants will be required to understand the mechanisms of action involved in growth responses and stress mitigation (Khan et al., 2009). The search for the mode of action of biostimulants is complicated by the observation that many biostimulants have been shown to induce genes and benefit productivity only when plants are challenged by abiotic and biotic stress. Experimental methods must therefore be developed to produce relevant and reproducible stress conditions so that the application of any molecular tool to probe gene function produces results that are relevant to the purported effects on plant productivity.

The role of signaling molecules in plant response to environmental cues has been an area of active research in plant biology. The process of signal transmission involves the synthesis of signaling molecules (ligands), their translocation, their binding to receptors, the resulting cellular responses, and, finally, the degradation of the signaling molecules (Zhao et al., 2005; Wang and Irving, 2011). When the signaling molecule binds to its receptor, the initial cellular response is the activation of secondary messengers, or intracellular signaling mediators, which cause a further series of cellular responses. Among the substances that may act as secondary messengers are: lipids, sugars, ions, nucleotides, gases, Ca^{2+} , cAMP, cGMP, cyclic ADP-ribose, small GTPase, 1,2-diacylglycerol, inositol-1,4,5-triphosphate, nitric oxide, phosphoinositides, and others (Zhao et al., 2005; Wang and Irving, 2011). Generally, a membrane-mediated action is typical for water-soluble compounds, while cytosol-mediated activity is primarily triggered by lipophilic compounds.

Whereas, enzymes interact with their substrates in a geometrical way (“lock and key”), signaling molecules are thought to have a topochemical affinity to their receptors.

It is assumed that the interaction of such components at the receptor site is cooperative and quantized (Gafurov and Zefirov, 2007). The bioactive compounds in some biostimulants are also proposed to display signaling activity in plants or induce signaling pathways. Various amino acids (Forde and Lea, 2007; Arbona et al., 2013), and peptides (Ivanov, 2010) function as signaling molecules in the regulation of plant growth and development (Ertani et al., 2009; Mochida and Shinozaki, 2011). Peptide signaling is important in various aspects of plant development and growth regulation including meristem organization, leaf morphogenesis, and defense responses to biotic and abiotic stress (Schiavon et al., 2008). Specific signaling peptides contained in a plant-derived protein hydrolysate have been shown to affect plant growth and development, defense responses, callus growth, meristem organization, root growth, leaf-shape regulation, and nodule development (Matsubayashi and Sakagami, 2006; Colla et al., 2013). Protein hydrolysates from soybean and casein have been shown to act as elicitors to enhance grapevine immunity against *Plasmopara viticola* (Lachhab et al., 2014).

Proteins may also contain hidden peptide sites, “cryptides” or “crypteins” in their amino acid sequence, which may have their own biological activities, distinct from its precursor (Ivanov, 2010; Samir and Link, 2011). Evidence that cryptides can trigger plants defense reactions have recently been demonstrated (Yamaguchi and Huffaker, 2011) and there are reports of the isolation of cryptides by hydrolysis of proteins from marine organisms, including seaweeds, and cryptides may be present naturally in a variety of biological derived products (Henda and Bordenave-Juchereau, 2014; Hayes et al., 2015).

Many small molecular weight substances are known to participate in signaling cascades *in vivo*. Exogenous amino acids may affect biological processes by acting directly as signal molecules or by influencing hormone action via amino acid conjugation (Tegeder, 2012). It has been suggested that amino acid based biostimulants are readily absorbed and translocated by plant tissues and once absorbed, they have the capacity to function as compatible osmolytes, transport regulators, signaling molecules, modulators of stomatal opening, and may detoxify heavy metals among other benefits (Kauffman et al., 2007). Sugars (Smeekens, 2000; Eveland and Jackson, 2012) and fatty acids and plant lipids (Kachroo and Kachroo, 2009) are also known to act as signaling molecules and mitigators of stress response in plants (Okazaki and Saito, 2014). Animal based lipid soluble fractions, have also been observed to produce an auxin-like response (Kauffman et al., 2007), while sugars, sucrose, and its cleavage products (hexoses), are also known to act as signaling molecules through regulation of gene expression and by interaction with other hormone signals including auxins. In a sunflower meal hydrolyzate, amino acids, humic substances, microelements, and sugars present in the biostimulant appeared to coordinate, with auxin-like compounds in complex signaling cross-talk promoting plant growth, enhancing plant transplanting success and increasing final crop yield (Ugolini et al., 2015).

Hormones are of central importance for the regulation of metabolic processes and plant development in a complex system of interacting hormones and cofactors, the functions of which

are closely intertwined and mutually dependent (Wang and Irving, 2011). Biostimulants developed from humic substances, complex organic materials, seaweeds, antitranspirants, free amino acids (Du Jardin, 2012), and crude extracts of lower (Rathore et al., 2009) and higher plants (Yakhin et al., 2012) have been frequently demonstrated to have an effect on plant hormonal status (Kurepin et al., 2014). While hormone-like compounds may be present in biostimulants, it is also possible that *de novo* synthesis of hormones may be induced by such preparations in treated plants (Jannin et al., 2012) and amino acids, glycosides, polysaccharides and organic acids are contained in many biostimulants and may act as precursors or activators of endogenous plant hormones (Paradić et al., 2011). Hormones or hormone-like effects could therefore be responsible for the action of natural biostimulants derived from microorganisms, algae, higher plants, animal, and humate based raw material (Table 4).

METABOLIC EFFECTS

Information on currently available biostimulants gives some insight into the possible biochemical and molecular genetic effects of biostimulants derived from different natural raw materials (Table 4). Many published reports are available suggesting various biostimulants improve plant productivity through increased assimilation of N, C, and S (Jannin et al., 2012, 2013), improved photosynthesis, improved stress responses, altered senescence, and enhanced ion transport (Gajic, 1989; Khan et al., 2009; Paradić et al., 2011). Biostimulants are also reported to increase free amino acids, protein, carbohydrates, phenolic compounds, pigment levels, and various enzymes (Table 4). The protective effect of many biostimulants against biotic and abiotic stresses has been associated with a reduction of stress-induced reactive oxygen species, activation of the antioxidant defense system of plants, or increased levels of phenolic compounds (Ertani et al., 2011a, 2013a).

While it is clear that many biologically derived biostimulants contain small molecular weight compounds that are involved in signaling events and may directly influence plant metabolic processes, it remains unclear how an exogenous soil or foliar application of an uncharacterized product can have predictable and beneficial responses in plants. It is well-known, for example, that application of exogenous plant hormones or compounds that disrupt hormone function (PGR's) can have markedly negative effects on plants and that optimization of PGR materials and their applications requires precise information on dosage and timing. Application of biostimulants for which the dosage and efficacy of the functional compounds is unknown, cannot, therefore, be expected to result in predictable plant responses and identification of molecules with effects on plant metabolic processes is not, in of itself, a sufficient explanation for the function of a biostimulant. It is also uncertain why the application of a biostimulant with purported function as a PGR, signaling molecule or other discrete compound would be superior to, or

more easily controlled, than a direct application of the purified product itself.

TOXICOLOGICAL AND ECOLOGICAL ASPECTS

Modern crop production requires a balance of high and consistent productivity with maximum safety for consumers, agricultural workers, and the environment (Rathore et al., 2009; Jannin et al., 2012; Pecha et al., 2012). While some biostimulants have been analyzed with regard to unwanted side effects including negative impact on the natural environment (Janas and Posmyk, 2013) most biostimulants have not been fully characterized but have been regarded as generally recognized as safe (GRAS in the US) on the basis of the biological origin of their constituents (Thomas et al., 2013). Generally, biostimulants are assumed to be biodegradable, non-toxic, non-polluting and non-hazardous to various organisms. While this may be a rational conclusion for many formulations derived from biological materials such as seaweed extracts and their components (Turan and Köse, 2004; Dhargalkar and Pereira, 2005; Rathore et al., 2009; Michalak and Chojnacka, 2014; Stadnik and de Freitas, 2014), higher plants (Onatsky et al., 2001; Abdalla, 2013; Yakhin et al., 2013), chitin and chitosan (Bautista-Baños et al., 2006; Cabrera et al., 2013) it is not clear that this is a valid assumption for microbial products or products that would not normally be present in agricultural fields.

Biostimulants have been utilized as bioremediants and have been shown to improve ATP levels and phosphatase and urease activity (Tejada et al., 2011a), and hence increase the rate of degradation of xenobiotics in the soil (Tejada et al., 2010, 2011b) and to enhance beneficial soil microbial communities under semi-arid climates (Tejada et al., 2011b). Biostimulants may also help reduce the amount of potentially risky agrochemicals (Kolomazník et al., 2012) including reducing the use of fertilizers and pesticides (Hamza and Suggars, 2001). Most compounds contained in biostimulants are natural constituents of terrestrial and aquatic ecosystems (Jannin et al., 2012) and metabolites of plant and microbial origin and as such most are generally regarded as safe, particularly at the low rates at which they are typically applied. Thus, it has been proposed that biostimulants can be positioned as eco-friendly products for sustainable agriculture (Mladenova et al., 1998; Ertani et al., 2011a; Ghannam et al., 2013; Vijayanand et al., 2014). In many countries, however, biostimulants are not subject to rigorous toxicological screening (Traon et al., 2014) and there remains the potential for the persistence of human pathogens in materials of animal origin and for the synthesis of novel compounds of unknown function or toxicology during the manufacturing process.

ECONOMIC ASPECTS

Even though there have been relatively few rigorous demonstrations of the benefit of biostimulants, and to a

large extent the mode of action of these products remains uncertain, the industry for biostimulants is substantial and rapidly growing. Though many recent “market” studies show that the market for these products is growing at a remarkable rate, the validity of these analyses must be considered with care as they frequently do not provide an explicit definition of term “biostimulants.” The value of the European biostimulants market ranged from €200 to €400 million in 2011, €500 million in 2013 and may grow to more than €800 million in 2018 with annual growth potential in 10% and more (EBIC, 2011a, 2013; Traon et al., 2014). France, Italy, Spain are the leading EU countries in the production of biostimulants (Traon et al., 2014). In North America, the biostimulant market was valued at \$0.27 billion in 2013¹, and is expected to grow at a growth rate of 12.4% annually, to reach \$0.69 billion by 2018, the USA is the largest producer and consumer of biostimulants in the region (<http://www.micromarketmonitor.com/>). In 2014, the USA market was assessed at \$313.0 million and is projected to reach \$605.1 million by 2019², at a CAGR of 14.1% (<http://news.agropages.com/>). The biostimulants market in the Asia-Pacific was valued at \$0.25 billion in 2013, and is expected to grow at a CAGR of 12.9% annually, to reach \$0.47 billion by 2018 (Asia Biostimulants Market, 2015)³. China and India are key countries playing a significant role. The Southeast Asian & Australasian biostimulants market was valued at \$233.8 million in 2015, and is projected to reach \$451.8 million by 2021 (<http://news.agropages.com/>)⁴. The market in Latin America was valued at \$0.16 billion in 2013⁵, and is expected to grow at a CAGR of 14.4% annually, to reach \$0.32 billion by 2018 (<http://www.micromarketmonitor.com/>). This market is mostly concentrated in Brazil and Argentina. The regional market shares of the global biostimulants market⁶ are: EU—41.7%, North America—21.5%, the Asia-Pacific region—20%, Latin America—12.9%. Globally, it biostimulants were valued at \$1402.15 million in 2014 and are projected to have aCAGR of 12.5% reaching \$2524.02 million by 2019⁷, largely as a consequence of growing interest in organic products. Wu (2016) summarised that “the global biostimulants market is projected to reach \$2.91 billion by 2021, with a CAGR (compound annual growth rate) of 10.4% from 2016 to 2021. In terms of area of application, the biostimulants market is projected to reach 24.9 million hectares by 2021 and is

¹North America biostimulants market (2015). Available online at <http://www.micromarketmonitor.com/> (Accessed February 27, 2015).

²North America biostimulants market to reach \$605.1 million by 2019 (2015). Available online at: <http://news.agropages.com/> (accessed August 18, 2015).

³Available online at: <http://www.micromarketmonitor.com/> (Accessed February 27, 2015).

⁴Southeast Asian and Australasian Biostimulants Market Trends and Forecasts to 2021 Available online at: <http://news.agropages.com/> (Accessed August 15, 2016).

⁵Latin America biostimulants market (2015). Available online at: <http://www.micromarketmonitor.com/> (Accessed February 27, 2015).

⁶Global biostimulants product market to reach \$2241.0 million by 2018. Available online at: <http://news.agropages.com/> (Accessed December 30, 2015).

⁷Global biostimulants market to reach \$2.52 bn by 2019. Available online at <http://news.agropages.com/> (Accessed December 30, 2015).

projected to grow at a CAGR of 11.7% from 2016 to 2021” (Wu, 2016).”

PROBLEMS AND PROSPECTS

The biostimulant industry faces many problems and challenges. Until recently biostimulant products based on natural raw materials and particularly waste stream has mainly been developed based on observational and less commonly, empirical data. While many contemporary biostimulants have been shown to be effective in practice, very few biostimulants can claim to understand the mechanisms or modes of action (Khan et al., 2009). Furthermore, while biostimulants can be categorized by source of origin, this is frequently inadequate as very substantial differences can exist between products even within a common feed stock origin. The challenge to biostimulant science is further exacerbated since composition and content of active substances in the original plant raw material can be affected by many factors including the location and growing conditions, season, species, variety, organ, and the phase of growth (Naumov et al., 1993; Dragovoz et al., 2009; Sharma et al., 2012b). Similarly, the response of the target crop can be expected to vary across crops and environments. One solution to this problem is to derive the raw materials for the biostimulant under highly regulated conditions. This approach has been successfully implemented by leading seaweed producers and fermentation based products that have developed harvesting and manufacturing processes that ensure uniformity of product performance through time. The development of a product with uniformity of response is not, however, a guarantee that the product is optimized for biological efficacy.

To address these issues, developments in -omics approaches will be critical in accelerating the discovery of mode of action of bioactive compounds (Aliferis and Jabaji, 2011; Craigie, 2011; Jannin et al., 2012) and optimizing their use. Metabolomics, phenomics and agronomics represent the integration of gene expression, protein interactions, and other regulatory processes as they impact on plant productivity and thus are more appropriate tools for discovery in this field than mRNA, transcripts, or proteins analyzed in isolation (Arbona et al., 2013). Integrative, multidisciplinary approaches using tools from transcriptomics in conjunction with metabolomics and biochemical analysis are necessary to establish the mechanism of action and to identify the active components in the extracts (Lee et al., 2012). The difficulty in identifying modes of action and subsequent standardization of composition of multicomponent biostimulants based on natural raw materials will continue to hamper the use, certification and registration of biostimulants. The solution to this problem will require the collaborative efforts of specialists from different fields: chemists, biologists, plant physiologists, industrial manufacture, sales and distribution and those with expertise in practical agricultural production (Raldugin, 2004; Craigie, 2011; Jannin et al., 2012; Lee et al., 2012).

Products with a single active substance represent a simpler construct in which the physiological effects and

mechanism of action can be more readily determined and hence certification and registration is simpler. The multicomponent composition of many preparations, however, are much more difficult to characterize (Bozhkov et al., 1996), though they may offer novel insight into biological synergy (Bulgari et al., 2015), multifunctionality and emergence which may be crucial to product efficacy (Gerhardson, 2002). In the absence of a functional rationale for every constituent in a multicomponent biostimulant, it is likely that there will be molecules present that may positively or negatively influence plant productivity. Currently, it is almost impossible using available chemical-synthetic, and genetic engineering approaches to reproduce the full suite of molecules and complexes of biologically active substances (Kershengolts et al., 2008) that are present in most biostimulants.

PRO'S AND CON'S OF BIOSTIMULANTS SCIENCE AND PRACTICE

Many have noted the state confusion in the field of biostimulants (Torre et al., 2013; Traon et al., 2014) and this has resulted in the opinion that much of the biostimulant market is not based on science or efficacy and that many products are little more than recycled waste products sold on the basis of pseudoscience and marketing. Indeed, research on several biostimulant products has shown them to be ineffective or to contain inactive, unstable or inconsistent properties with several showing negative effects compared when contrasted with well-designed controls (Csizinszky, 1984, 1986; Albrechts et al., 1988; Di Marco and Osti, 2009; Vasconcelos et al., 2009; Banks and Percival, 2012; Cerdan et al., 2013; de Oliveira et al., 2013; Carvalho et al., 2014). For example, foliar and root application of a product containing amino acids from animal origin have been reported to cause severe plant-growth depression and negative effects on Fe nutrition while a second product containing amino acids from plant origin stimulated plant growth (Cerdan et al., 2013). In another report that tested several biostimulant products it was concluded that “none of the biostimulant products tested achieved a sufficient degree of pathogen control to warrant replacement of or supplementation with conventional synthetic fungicides” (Banks and Percival, 2012), and there have been demonstrated positive and negative impacts and overall questions of the economic feasibility of the use of humic substances for increasing crop yields (Rose et al., 2014). Since biological systems are inherently complex, and given that most biostimulant products have not been characterized and have received relatively little replicated and rigorous independent validation, it is perhaps not surprising that many products are ineffective or highly variable in response. Nevertheless, there are a significant number of rigorous independent reports of benefits from some biostimulant formulations and market growth data demonstrates that there is a good deal of support for these products within agricultural producer communities. That such market growth has occurred, even in the absence of a known “mechanism of function” suggests that there are aspects of plant

metabolism and productivity constraints that are not understood but are potentially important if we are to achieve the goal of increased global food production.

The market euphoria that is taking place in the biostimulant industry recognizes these unknowns and biostimulants are viewed by many innovators and investors as a mechanism to conduct broadscale, if unfocused, discovery of novel biologically derived molecules. Much as the exploration of marine organisms, and plants and microbes from diverse ecosystems has led to the discovery of novel pharmaceuticals, so too the development of biostimulants from the broad range of source materials, holds significant promise of discovery. Recent years have seen rapid growth in the number of published studies, increased numbers of scientific conferences and development of legal framework and legislation. These trends will inevitably improve the image of this industry and the efficacy of products. Two significant problems still exist within the industry broadly: (1) preparations of products with highly complex multicomponent and incompletely identified composition make the identification of a primary mode of action extremely difficult and (2) the current classification and legislation/legal framework for regulation of biostimulants is based primarily on source material and not on biological mode of action. Hence there is insufficient capacity to differentiate products, and there is the potential for the successful demonstration of a single product within a biostimulant category, to falsely indicate the efficacy of the group as whole.

Several topical questions need consideration in the future:

1. Can living cultures of microorganisms, which have the ability to stimulate the growth of plants be referred to biostimulants?
2. Are non-essential elements that result in improved plant productivity, biostimulants?
3. How should biostimulants with a complex completely unidentified structure where all the components and modes/mechanisms involved have not been established be registered and regulated in national and international legislation?
4. What standard of proof of efficacy is appropriate that both stimulates development and discourages the sale of materials of no benefit?
5. On what principles, should the final classification of biostimulants be based and what categories should it contain?

CONCLUSIONS

Modern biostimulants are complex mixtures derived from raw materials of highly diverse origin utilizing highly diverse manufacturing processes and as such can be expected to have a broad spectrum of possible biological activity and safety. To distinguish biostimulants from the existing legislative product categories including essential nutrients, pesticides, or plant hormones a biostimulant should not solely function by virtue of the presence of elements or compounds of known function. We propose, therefore, a definition of a biostimulant as “a formulated product of biological origin that improves plant productivity as a consequence of the novel or emergent properties of the complex

of constituents and not as a sole consequence of the presence of known essential plant nutrients, plant growth regulators, or plant protective compounds.” Consistent with this definition, the ultimate identification of a novel molecule within a biostimulant that is found to be wholly responsible for the biological function of that biostimulant, would necessitate the classification of the biostimulant according to the discovered function.

This novel definition is inspired by three observations: (1) that the development of the biostimulant industry will inevitably result in the discovery of novel biologically active molecules and that the identification and classification of these molecules will benefit biological discovery more greatly if these molecules are explicitly described than if they were merely labeled as “biostimulants,” (2) that there is a need for the nascent biostimulant industry to explicitly discourage the inclusion of nutrient elements and known biologically active molecules under the guise of a “biostimulant” and (3) that there is a need to recognize that classic reductionist biology/chemistry may indeed be insufficient to explain biological complexity (Luisi, 2002; Lüttge, 2012; Bertolli et al., 2014).

The definition provided here is important as it emphasizes the principle that biological function can be modulated through application of complex mixtures of molecules for which an explicit mode of action has not been defined. The definition also requires a demonstration of beneficial impacts of the biostimulant on plant productivity. Given the difficulty in determining a “mode of action” for a biostimulant, and recognizing the need for the market in biostimulants to attain legitimacy, we suggest that the focus of biostimulant research and validation should be upon determining the mechanism of action, without a requirement for the determination of a mode of action. This can be achieved through careful agronomic experimentation, molecular or biochemical demonstration of positive impact on biological processes or the use of advanced analytical equipment to identify functional constituents. Given the prerequisite multi-component and emergent characteristics of biostimulants, the discovery of the mode of action is likely to require application of new techniques in bioinformatics and systems biology. While the definition proposed here suggests that the development and marketing of a biostimulant does not require a demonstration of the mode of action, it is still in the interest of the commercial producers of these products to pursue an understanding of these products so that the product can be improved and optimized for use in various environments and cropping systems.

While there is a clear commercial imperative to rationalize biostimulants as a discrete class of products, there is also a compelling biological case for the science-based development of the biostimulant science that is grounded in the observation that the application of biological materials derived from various organisms, including plants, that have been exposed to stressors can affect metabolic and energetic processes in humans, animals, and plants (Filatov, 1951a,b). This hypothesis is based upon the premise that functional chemical communication occurs between individuals or organs that favorably modulate metabolic pathways and networks at different plant hierarchical levels. Inter and intra organism communication and consequent molecular

and metabolic regulation are at the heart of the science of systems biology and the tools of systems biology will inevitably be critical to the realization of mode of action of many biostimulants. Continued investments by commercial entities in biostimulant research and product development will serve as a critical driver of discovery in this realm and will inevitably lead to the identification of novel biological phenomenon, pathways and processes that would not have been discovered if the category of biostimulants did not exist, or was not considered legitimate.

REFERENCES

- Abbas, S. M. (2013). The influence of biostimulants on the growth and on the biochemical composition of *Vicia faba* CV. Giza 3 beans. *Rom. Biotech. Lett.* 18, 8061–8068. Available online at: <http://www.rombio.eu/vol18nr2/1%20Salwa%20Mohamed.pdf>
- Abd El-Baky, H. H., Hussein, M. M., and El-Baroty, G. S. (2008). Algal extracts improve antioxidant defense abilities and salt tolerance of wheat plant irrigated with sea water. *Afr. J. Biochem. Res.* 7, 151–164. Available online at: <http://www.academicjournals.org/journal/AJBR/article-abstract/82070DD10085>
- Abdalla, M. M. (2013). The potential of *Moringa oleifera* extract as a biostimulant in enhancing the growth, biochemical and hormonal contents in rocket (*Eruca vesicaria subsp. sativa*) plants. *Int. J. Plant Physiol. Biochem.* 5, 42–49. doi: 10.5897/IJPPB2012.026
- Adani, F., Genevini, P., Zaccheo, P., and Zocchi, G. (1998). The effect of commercial humic acid on tomato plant growth and mineral nutrition. *J. Plant. Nutr.* 21, 561–575. doi: 10.1080/01904169809365424
- Adholeya, A., Tiwari, P., and Singh, R. (2005). “Large-scale production of arbuscular mycorrhizal fungi on root organs and inoculation strategies,” in *In vitro Culture of Mycorrhizas*, eds S. Declerck, D. G. Strullu, and J. A. Fortin (Heidelberg: Springer), 315–338.
- Aguado-Santacruz, G. A., Moreno-Gómez, B., Rascón-Cruz, Q., Aguirre-Mancilla, C., Espinosa-Solis, J. A., and González-Barriga, C. D. (2014). “Biofertilizers as complements to synthetic and organic fertilization,” in *Components, Uses in Agriculture and Environmental Impacts*, eds F. López-Valdes and F. Fernández-Luqueno (New York, NY: Nova Science Publishers Inc.), 155–180.
- Aguirre, E., Leménager, D., Bacaicoa, E., Fuentes, M., Baigorri, R., Angel, M. Z., et al. (2009). The root application of a purified leonardite humic acid modifies the transcriptional regulation of the main physiological root responses to Fe deficiency in Fe-sufficient cucumber plants. *Plant Physiol. Biochem.* 47, 215–223. doi: 10.1016/j.plaphy.2009.06.004
- Aharoni, A., and Galili, G. (2011). Metabolic engineering of the plant primary-secondary metabolism interface. *Curr. Opin. Biotechnol.* 22, 239–244. doi: 10.1016/j.copbio.2010.11.004
- Aitken, J. B., and Senn, T. L. (1965). Seaweed products as a fertilizer and soil conditioner for horticultural crops. *Bot. Mar.* 8, 144–148. doi: 10.1515/botm.1965.8.1.144
- Albregts, E. E., Howard, C. M., Chandler, C., and Mitchell, R. L. (1988). Effect of biostimulants on fruiting of strawberry. *Proc. Fla. State Hort. Soc.* 101, 370–372.
- Aliferis, K. A., and Jabaji, S. (2011). Metabolomics – a robust bioanalytical approach for the discovery of the modes-of-action of pesticides: a review. *Pestic. Biochem. Phys.* 100, 105–117. doi: 10.1016/j.pestbp.2011.03.004
- Apone, F., Arciello, S., Colucci, G., Filippini, L., and Portoso, D. (2006). Alle radici della biostimolazione: indagini scientifiche a supporto. *Fertilitas Agrorum* 1, 55–63. Available online at: http://fertilitasagrorum.ciec-italia.it/index_file/volumi.htm
- Apone, F., Tito, A., Carola, A., Arciello, S., Tortora, A., Filippini, L., et al. (2010). A mixture of peptides and sugars derived from plant cell walls increases plant defense responses to stress and attenuates ageing-associated molecular changes in cultured skin cells. *J. Biotechnol.* 145, 367–376. doi: 10.1016/j.jbiotec.2009.11.021
- Arbona, V., Manzi, M., Ollas, C., and Gómez-Cadenas, A. (2013). Metabolomics as a tool to investigate abiotic stress tolerance in plants. *Int. J. Mol. Sci.* 14, 4885–4911. doi: 10.3390/ijms14034885

AUTHOR CONTRIBUTIONS

All authors OY, AL, IY, PB, contributed equally to this review.

ACKNOWLEDGMENTS

We thank Russian Foundation for Basic Research (RFBR-Agidel No 02-04-97917), Russian Science Support Foundation, Foundation for Assistance to Small Innovative Enterprises in Science and Technology (“Start Program,” FASIE, Russia).

- Aremu, A. O., Masondo, N. A., Rengasamy, K. R. R., Amoo, S. O., Gruz, J., Bība, O., et al. (2015b). Physiological role of phenolic biostimulants isolated from brown seaweed *Ecklonia maxima* on plant growth and development. *Planta* 246, 1313–1324. doi: 10.1007/s00425-015-2256-x
- Aremu, A. O., Stirk, W. A., Kulkarni, M. G., Tarkowská, D., Turečková, V., Gruz, J., et al. (2015a). Evidence of phytohormones and phenolic acids variability in garden-waste-derived vermicompost leachate, a well-known plant growth stimulant. *Plant Growth Regul.* 75, 483–492. doi: 10.1007/s10725-014-0011-0
- Arnao, M. B., and Hernández-Ruiz, J. (2014). Melatonin: plant growth regulator and/or biostimulator during stress? *Trends Plant Sci.* 19, 789–797. doi: 10.1016/j.tplants.2014.07.006
- Arthur, G. D., Aremu, A. O., Moyo, M., Stirk, W. A., and van Staden, J. (2013). Growth-promoting effects of a seaweed concentrate at various pH and water hardness conditions. *S. Afr. J. Sci.* 109, 1–6. doi: 10.1590/sajs.2013/20120013
- Aydin, A., Kant, C., and Turan, M. (2012). Humic acid application alleviate salinity stress of bean (*Phaseolus vulgaris* L.) plants decreasing membrane leakage. *Afr. J. Agric. Res.* 7, 1073–1086. doi: 10.5897/ajar10.274
- Aylward, L. (2005). More superintendents are using biostimulants and trusting in them, companies say. *Golfdom* 54–58. doi: 10.5897/AJAR10.274
- Baglieri, A., Cadili, V., Mozzetti Monterumici, C., Gennari, M., Tabasso, S., Montoneri, E., et al. (2014). Fertilization of bean plants with tomato plants hydrolysates. Effect on biomass production, chlorophyll content and N assimilation. *Sci. Hortic.* 176, 194–199. doi: 10.1016/j.scienta.2014.07.002
- Banks, J., and Percival, G. C. (2012). Evaluation of biostimulants to control Guignardia leaf blotch (*Guignardia aesculi*) of horsechestnut and black spot (*Diplocarpon rosae*) of roses. *Arboric. Urban Forest.* 38, 258–261. Available online at: <http://www.ncufc.org/uploads/biostimulantsanddisease.pdf>
- Bargiacchi, E., Miele, S., Romani, A., and Campo, M. (2013). Biostimulant activity of hydrolyzable tannins from sweet chestnut (*Castanea sativa* Mill.). *Acta Hortic.* 1009, 111–116. doi: 10.17660/ActaHortic.2013.1009.13
- Basak, A. (2008). “Biostimulators – definitions, classification and legislation,” in *Monographs Series: Biostimulators in Modern Agriculture. General Aspects*, ed H. Gawrońska (Warsaw: Wiesz Jutra), 7–17.
- Bashan, Y. (1998). Inoculants of plant growth-promoting bacteria for use in agriculture. *Biotechnol. Adv.* 16, 729–770. doi: 10.1016/S0734-9750(98)00003-2
- Bautista-Baños, S., Hernández-Lauzardo, A. N., Velázquez-del Valle, M. G., Hernández-López, M., Ait Barka, E., Bosquez-Molina, E., et al. (2006). Chitosan as a potential natural compound to control pre and postharvest diseases of horticultural commodities. *Crop Prot.* 25, 108–118. doi: 10.1016/j.cropro.2005.03.010
- Beaudreau, D. G. Jr. (2013). *Biostimulants in Agriculture: Their Current and Future Role in a Connected Agricultural Economy*. Available online at: <http://www.biostimulantscoalition.org/> (Accessed September 27, 2014).
- Belakbir, A., Ruiz, J. M., and Romero, L. (1998). Yield and fruit quality of pepper (*Capsicum annuum* L.) in response to bioregulators. *HortScience*, 33, 85–87.
- Berbara, R. L. L., and Garcia, A. C. (2014). “Humic substances and plant defense metabolism,” in *Physiological Mechanisms and Adaptation Strategies in Plants Under Changing Environment*, eds P. Ahmad and M. R. Wani (New York, NY: Springer Science+Business Media), 297–319.
- Bertolli, S. C., Mazzafera, P., and Souza, G. M. (2014). Why is it so difficult to identify a single indicator of water stress in plants? a proposal for a multivariate analysis to assess emergent properties. *Plant Biol.* 16, 578–585. doi: 10.1111/plb.12088

- Betti, L., Canova, A., Maini, P., Merendino, A., and Paolini, M. (1992). Effects of foliar application of an aminoacid-based biostimulant on the response of pepper seedling to PepMV infections. *Adv. Hortic. Sci.* 6, 97–103.
- Bhalla, U. S., and Iyengar, R. (1999). Emergent properties of networks of biological signaling pathways. *Science* 283, 381–387. doi: 10.1126/science.283.5400.381
- Bhardwaj, D., Ansari, M. W., Sahoo, R. K., and Tuteja, N. (2014). Biofertilizers function as key player in sustainable agriculture by improving soil fertility, plant tolerance and crop productivity. *Microb. Cell Fact.* 13:66. doi: 10.1186/1475-2859-13-66
- Bhattacharyya, P. N., and Jha, D. K. (2012). Plant growth promoting rhizobacteria (PGPR): emergence in agriculture. *World J. Microbiol. Biotechnol.* 28, 1327–1350. doi: 10.1007/s11274-011-0979-9
- Billard, V., Etienne, P., Jannin, L., Garnica, M., Cruz, F., Garcia-Mina, J.-M., et al. (2014). Two biostimulants derived from algae or humic acid induce similar responses in the mineral content and gene expression of winter oilseed rape (*Brassica napus* L.). *J. Plant Growth Regul.* 33, 305–316. doi: 10.1007/s00344-013-9372-2
- Blagoveshchensky, A. V. (1945). Biochemical factors of natural selection in plants. *J. Gen. Biol.* 6, 217–234.
- Blagoveshchensky, A. V. (1955). Biogenic stimulants in agriculture. *Priroda* 7, 43–47.
- Blagoveshchensky, A. V. (1956). Biogenic stimulants and biochemical nature of their action. *Bull. Main Bot. Garden.* 25, 79–86.
- Borri, R. (2011). “Use of plant-associated *Bacillus* strains as biofertilizers and biocontrol agents in agriculture,” in *Bacteria in Agrobiology: Plant Growth Responses*, ed D. K. Maheshwari (Berlin: Springer), 41–76.
- Botta, A., Sierras, N., Sánchez-Hernández, L., Martínez, M. J., and Bru, R. (2015). “Understanding the effect of an amino acids based biostimulant by an enantiomeric analysis of their active principles and a proteomic profiling approach,” in *Abstracts Book for Oral and Poster Presentations of the 2st World Congress on the use of Biostimulants in Agriculture; 2015 Nov 16-19*, eds P. Perata, P. Brown, R. A. Alvarez, and M. Ponchet (Florence: New Ag International), 40.
- Bozhkov, A. I., Menzyanova, N. G., and Leontovich, V. P. (1996). Lipid-composition and antibacterial activity of root exudates secreted by wheat seedlings. *Russ. J. Plant Physiol.* 43, 795–799.
- Brice-o-Domínguez, D., Hernández-Carmona, G., Moyo, M., Stirk, W., and van Staden, J. (2014). Plant growth promoting activity of seaweed liquid extracts produced from *Macrocystis pyrifera* under different pH and temperature conditions. *J. Appl. Phycol.* 26, 2203–2210. doi: 10.1007/s10811-014-0237-2
- Brown, P., and Saa, S. (2015). Biostimulants in agriculture. *Front. Plant Sci.* 6:671. doi: 10.3389/fpls.2015.00671
- Bulgari, R., Cocetta, G., Trivellini, A., Vernieri, P., and Ferrante, A. (2015). Biostimulants and crop responses: a review. *Biol. Agric. Hortic.* 31, 1–17. doi: 10.1080/01448765.2014.964649
- Cabrera, J. C., Wégria, G., Onderwater, R. C. A., Nápoles, M. C., Falcón-Rodríguez, A. B., Costales, D., et al. (2013). Practical use of oligosaccharins in agriculture. *Acta Hortic.* 1009, 195–212. doi: 10.17660/ActaHortic.2013.1009.24
- Cacco, G., and Dell’Agnola, G. (1984). Plant growth regulator activity of soluble humic complexes. *Can. J. Soil Sci.* 64, 225–228. doi: 10.4141/cjss84-023
- Calvo, P., Nelson, L., and Kloepper, J. W. (2014). Agricultural uses of plant biostimulants. *Plant Soil* 383, 3–41. doi: 10.1007/s11104-014-2131-8
- Cambri, D., Filippini, L., Apone, F., Arciello, S., Colucci, G., and Portoso, D. (2008). “Effect of aminoplant® on expression of selected genes in *Arabidopsis thaliana* L. plants,” in *Biostimulants in Modern Agriculture. General Aspects*, ed H. Gawrońska (Warsaw: Wieś Jutra), 77–82.
- Canellas, L. P., and Olivares, F. L. (2014). Physiological responses to humic substances as plant growth promoter. *Chem. Biol. Technol. Agric.* 1:3. doi: 10.1186/2196-5641-1-3
- Canellas, L. P., Olivares, F. L., Okorokova-Façanha, A. L., and Façanha, A. R. (2002). Humic acids isolated from earthworm compost enhance root elongation, lateral root emergence, and plasma membrane H⁺-ATPase activity in maize roots. *Plant Physiol.* 130, 1951–1957. doi: 10.1104/pp.007088
- Canellas, L. P., Piccolo, A., Dobbss, L. B., Spaccini, R., Olivares, F. L., Zandonadi, D. B., et al. (2010). Chemical composition and bioactivity properties of size-fractions separated from a vermicompost humic acid. *Chemosphere* 78, 457–466. doi: 10.1016/j.chemosphere.2009.10.018
- Carvalho, M. E. A., Castro, P. R. D. C. E., Gallo, L. A., and Ferraz, M. V. D. C. Jr. (2014). Seaweed extract provides development and production of wheat. *Rev. Agrarian* 7, 166–170. doi: 10.5281/zenodo.51607
- Caulet, R. P., Gradinariu, G., Iurea, D., and Morariu, A. (2014). Influence of furostanol glycosides treatments on strawberry (*Fragaria × ananassa* Duch.) growth and photosynthetic characteristics under drought condition. *Sci. Hortic.* 169, 179–188. doi: 10.1016/j.scienta.2014.02.031
- Cerdan, M., Sanchez-Sanchez, A., Jorda, J. D., Juarez, M., and Sanchez-Andreu, J. (2013). Effect of commercial amino acids on iron nutrition of tomato plants grown under lime-induced iron deficiency. *J. Plant Nutr. Soil Sci.* 176, 859–866. doi: 10.1002/jpln.201200525
- Chambers, J. W. (2014). *Tea Extracts and Uses in Promoting Plant Growth*. U.S. Patent No. 20140113814 A1, 27. Available online at: <http://www.freepatentsonline.com/y2014/0113814.html>
- Chambolle, C. (2005). Biostimulants: humus substances. *PHM Rev. Hortic.* 468, 21–23. Available online at: <https://www.cabdirect.org/cabdirect/abstract/20053088318>
- Chbani, A., Mawlawi, H., and Zaouk, L. (2013). Evaluation of brown seaweed (*Padina pavonica*) as biostimulant of plant growth and development. *Afr. J. Agric. Res.* 13, 1155–1165. doi: 10.5897/AJAR12.1346
- Chen, T., Gu, J., Zhang, X., Ma, Y., Cao, L., Wang, Z., et al. (2014). System-level study on synergism and antagonism of active ingredients in traditional chinese medicine by using molecular imprinting technology. *Sci. Rep.* 4:7159. doi: 10.1038/srep07159
- Chen, Y., Clapp, C. E., and Magen, H. (2004). Mechanisms of plant growth stimulation by humic substances: the role of organo iron complexes. *Soil Sci. Plant Nutr.* 50, 1089–1095. doi: 10.1080/00380768.2004.10408579
- Chiu, H. C., Levy, R., and Borenstein, E. (2014). Emergent biosynthetic capacity in simple microbial communities. *PLoS Comput. Biol.* 10:e1003695. doi: 10.1371/journal.pcbi.1003695
- Chojnacka, K., Michalak, I., Dmytryk, A., Wilk, R., and Gorecki, H. (2015). “Innovative natural plant growth biostimulants,” in *Fertilizer Technology: II Biofertilizer*, eds S. Sinha, K. K. Pant, S. Bajpai, and J. N. Govil (Houston, TX: Studium Press LLC), 451–489.
- Christofolletti, C. A., Escher, J. P., Correia, J. E., Marinho, J. F. U., and Fontanetti, C. S. (2013). Sugarcane vinasse: environmental implications of its use. *Waste Manage.* 33, 2752–2761. doi: 10.1016/j.wasman.2013.09.005
- Ciavatta, C., and Cavani, L. (2006). Problematiche per l’inserimento dei biostimolanti nella legislazione dei fertilizzanti. *Fertilitas Agrorum* 1, 11–15. Available online at: http://fertilitasagrorum.ciec-italia.it/index_file/volumi.htm
- Ciesiolka, D., Gulewicz, P., Martinez-Villaluenga, C., Pilarski, R., Bednarczyk, M., and Gulewicz, K. (2005). Products and biopreparations from alkaloid-rich lupin in animal nutrition and ecological agriculture. *Folia Biol.* 53, 59–66. doi: 10.3409/173491605775789443
- Colla, G., Rouphael, Y., Canaguier, R., Svecova, E., and Cardarelli, M. (2014). Biostimulant action of a plant-derived protein hydrolysate produced through enzymatic hydrolysis. *Front. Plant Sci.* 5:448. doi: 10.3389/fpls.2014.00448
- Colla, G., Rouphael, Y., Di Mattia, E., El-Nakhel, C., and Cardarelli, M. (2015). Co-inoculation of *Glomus intraradices* and *Trichoderma atroviride* acts as a biostimulant to promote growth, yield and nutrient uptake of vegetable crops. *J. Sci. Food Agric.* 95, 1706–1715. doi: 10.1002/jsfa.6875
- Colla, G., Svecova, E., Rouphael, Y., Cardarelli, M., Reynaud, H., Canaguier, R., et al. (2013). Effectiveness of a plant-derived protein Hydrolysate to improve crop performances under different growing conditions. *Acta Hortic.* 1009, 175–180. doi: 10.17660/ActaHortic.2013.1009.21
- Conan, C., Guiboileau, A., Joubert, J.-M., and Potin, P. (2015). “Investigations of seaweed filtrate as Biostimulant,” in *Abstracts Book for Oral and Poster Presentations of the 2st World Congress on the use of Biostimulants in Agriculture; 2015 Nov 16-19*, eds P. Perata, P. Brown, R. A. Alvarez, and M. Ponchet (Florence: New Ag International), 75.
- Cordeiro, F. C., Santa-Catarina, C., Silveira, V., and Souza, S. R. D. (2011). Humic acid effect on catalase activity and the generation of reactive oxygen species in corn (*Zea mays*). *Biosci. Biotechnol. Biochem.* 75, 70–74. doi: 10.1271/bbb.100553
- Corte, L., Dell’Abate, M. T., Magini, A., Migliore, M., Felici, B., Roscini, L., et al. (2014). Assessment of safety and efficiency of nitrogen organic fertilizers from

- animal-based protein hydrolysates—a laboratory multidisciplinary approach. *J. Sci. Food Agr.* 94, 235–245. doi: 10.1002/jsfa.6239
- Craigie, J. S. (2011). Seaweed extract stimuli in plant science and agriculture. *J. Appl. Phycol.* 23, 371–393. doi: 10.1007/s10811-010-9560-4
- Craigie, J. S., MacKinnon, S. L., and Walter, J. A. (2009). Liquid seaweed extracts identified using ¹H NMR profiles. *J. Appl. Phycol.* 20, 665–671. doi: 10.1007/s10811-007-9232-1
- Crouch, I. J., and van Staden, J. (1993a). Commercial seaweed products as biostimulants in horticulture. *J. Home Consum. Hort.* 1, 19–76.
- Crouch, I. J., and van Staden, J. (1993b). Evidence for the presence of plant growth regulators in commercial seaweed products. *Plant Growth Reg.* 13, 21–29.
- Crouch, I. J., Smith, M. T., van Staden, J., Lewis, M. J., and Hoad, G. V. (1992). Identification of auxins in a commercial seaweed concentrate. *J. Plant. Physiol.* 139, 590–594. doi: 10.1016/S0176-1617(11)80375-5
- Csizinszky, A. A. (1984). Response of tomatoes to seaweed based nutrient sprays. *Proc. Fla. State Hort. Soc.* 97, 151–157.
- Csizinszky, A. A. (1986). Response of tomatoes to foliar biostimulant sprays. *Proc. Fla. State Hort. Soc.* 99, 353–358.
- Cutler, H. G., and Cutler, S. J. (2004). “Growth regulators, plant,” in *Kirk-Othmer Encyclopedia of Chemical Technology*. John Wiley and Sons Inc. doi: 10.1002/0471238961.1612011403212012.a01.pub2
- Daniels, R. S. (2013). *Corn Steep Liquor as A Biostimulant Composition*. U.S. Patent No. 8568758 B2, 12. Available online at: <http://www.freepatentsonline.com/y2012/0028801.html>
- de Fretes, C. E., Sembiring, L., and Purwestri, Y. A. (2013). Characterization of *Streptomyces* spp. producing indole-3-acetic acid as biostimulant agent. *Indones. J. Biotech.* 18, 83–91. doi: 10.22146/ijbiotech.7872
- De Lucia, B., and Vecchiatti, L. (2012). Type of bio-stimulant and application method effects on stem quality and root system growth in L.A. Lily. *Europ. J. Hort. Sci.* 77, 10–15. Available online at: <http://www.jstor.org/stable/24126519>
- de Oliveira, F. D. A. D., Medeiros, J. F. D., Oliveira, M. K. T. D., Souza, A. A. T., Ferreira, J. A., and Souza, M. S. (2013). Interaction between water salinity and biostimulant in the cowpea plants. *Rev. Brasil. de Eng. Agríc. Ambien.* 17, 465–471. doi: 10.1590/S1415-43662013000500001
- Dhargalkar, V. K., and Pereira, N. (2005). Seaweed: promising plant of the millennium. *Sci. Cult.* 71, 60–66. Available online at: <http://drs.nio.org/drs/handle/2264/489>
- Di Marco, S., and Osti, F. (2009). Effect of biostimulant sprays on *Phaeoconiella chlamydospora* and esca proper infected vines under greenhouse and field conditions. *Phytopathol. Mediterr.* 48, 47–58. doi: 10.14601/Phytopathol_Mediterr-2874
- Dixon, G. R., and Walsh, U. F. (2004). “Suppressing *Pythium ultimum* induced damping-off in cabbage seedlings by biostimulation with proprietary liquid seaweed extracts managing soil-borne pathogens: a sound rhizosphere to improve productivity in intensive horticultural systems,” in *Proceedings of the XXVI Inter. Horticultural Congr* (Toronto, ON).
- Doak, S. O., Schmidt, R. E., and Ervin, E. H. (2005). Metabolic enhancer impact on creeping bentgrass leaf sodium and physiology under salinity. *Int. Turfgr. Soc. Res. J.* 10, 845–849. Available online at: <http://turfgrassociety.com/itsjournal.html>
- Dobbss, L. B., Canellas, L. P., Olivares, F. L., Aguiar, N. O., Peres, L. E. P., Azevedo, M., et al. (2010). Bioactivity of chemically transformed humic matter from vermicompost on plant root growth. *J. Agric. Food Chem.* 58, 3681–3688. doi: 10.1021/jf904385c
- Dragovoz, I. V., Yavorskaya, V. K., Antoniuk, V. P., and Kurchii, B. A. (2009). Hormonal substances produced by microorganism association from ginseng roots. *Physiol. Biochem. Cultivated plants* 41, 393–399.
- Dubravac, K., Dubravac, I., and Maniasevic, J. (1995). The effect of the bioregulators Agrispin® and Ergostim on the vegetative and reproductive growth of apples. *J. Sustain. Agr.* 5, 73–83. doi: 10.1300/J064v05n01_06
- Du Jardin, P. (2012). *The Science of Plant Biostimulants - A Bibliographic Analysis, Ad hoc Study Report*. Brussels: European Commission. Available online at: <http://hdl.handle.net/2268/169257> (Accessed April 25, 2013).
- Du Jardin, P. (2015). Plant biostimulants: definition, concept, main categories and regulation. *Sci. Hortic.* 196, 3–14. doi: 10.1016/j.scienta.2015.09.021
- Dumas, B., Vergnes, S., Attia, F., and Noël, D. (2012). “Characterization of a new phyto-stimulating preparation: mode of action and evaluation of agronomic performance,” in *Abstracts Book for Oral and Poster Presentations of the 1st World Congress on the use of Biostimulants in Agriculture; 2012 Nov 26-29*, eds P. Perata, P. Brown, and M. Ponchet (Strasbourg: New Ag International), 86.
- EBIC (2011a). *Economic Overview of the Biostimulants Sector in Europe*. Available online at: <http://www.biostimulants.eu/> (Accessed October 23, 2012).
- EBIC (2011b). Available online at: <http://www.biostimulants.eu/2011/10/biostimulants-definition-agreed/> (Accessed July 29, 2014)
- EBIC (2012). Available online at: <http://www.biostimulants.eu/> (Accessed September 27, 2014).
- EBIC (2013). *Economic Overview of the Biostimulants Sector in Europe*. Available online at: <http://www.biostimulants.eu/> (Accessed September 23, 2014).
- Elliott, M. L., and Prevatte, M. (1996). Response of ‘Tifdwarf’ Bermudagrass to Seaweed-derived Biostimulants. *HortTechnology* 6, 261–263.
- Ertani, A., Cavani, L., Pizzeghello, D., Brandellero, E., Altissimo, A., Ciavatta, C., et al. (2009). Biostimulant activity of two protein hydrolysates in the growth and nitrogen metabolism of maize seedlings. *J. Plant Nutr. Soil Sci.* 172, 237–244. doi: 10.1002/jpln.200800174
- Ertani, A., Francioso, O., Tugnoli, V., Righi, V., and Nardi, S. (2011b). Effect of commercial liginosulfonate-humate on *Zea mays* L. metabolism. *J. Agric. Food Chem.* 59, 11940–11948. doi: 10.1021/jf202473e
- Ertani, A., Pizzeghello, D., Altissimo, A., and Nardi, S. (2013b). Use of meat hydrolysate derived from tanning residues as plant biostimulant for hydroponically grown maize. *J. Plant Nutr. Soil Sci.* 176, 287–295. doi: 10.1002/jpln.201200020
- Ertani, A., Pizzeghello, D., Baglieri, A., Cadili, V., Tambone, F., Gennari, M., et al. (2013c). Humic-like substances from agro-industrial residues affect growth and nitrogen assimilation in maize (*Zea mays* L.) plantlets. *J. Geochem. Explor.* 129, 103–111. doi: 10.1016/j.jgexplo.2012.10.001
- Ertani, A., Pizzeghello, D., Francioso, O., Sambo, P., Sanchez-Cortes, S., and Nardi, S. (2014). *Capsicum chinensis* L. growth and nutraceutical properties are enhanced by biostimulants in a long-term period: chemical and metabolomic approaches. *Front. Plant Sci.* 5:375. doi: 10.3389/fpls.2014.00375
- Ertani, A., Schiavon, M., Altissimo, A., Franceschi, C., and Nardi, S. (2011a). Phenol-containing organic substances stimulate phenylpropanoid metabolism in *Zea mays* L. *J. Plant Nutr. Soil Sci.* 174, 496–503. doi: 10.1002/jpln.201000075
- Ertani, A., Schiavon, M., Muscolo, A., and Nardi, S. (2013a). Alfalfa plant-derived biostimulant stimulate short-term growth of salt stressed *Zea mays* L. plants. *Plant Soil* 364, 145–158. doi: 10.1007/s11104-012-1335-z
- Eveland, A. L., and Jackson, D. P. (2012). Sugars, signalling, and plant development. *J. Exp. Bot.* 63, 3367–3377. doi: 10.1093/jxb/err379
- Featonby-Smith, B. C., and van Staden, J. (1983). The effect of seaweed concentrate on the growth of tomato plants in nematode-infested soil. *Sci. Hortic.* 20, 137–146. doi: 10.1016/0304-4238(83)90134-6
- Ferrante, A., Trivellini, A., Vernieri, P., and Piaggini, A. (2013). Application of Actiwave® for improving the rooting of Camellia cuttings. *Acta Hortic.* 1009, 213–218. doi: 10.17660/ActaHortic.2013.1009.25
- Filatov, V. P. (1944). Tissue therapy in ophthalmology. *Am. Rev. Sov. Med.* 2, 53–66.
- Filatov, V. P. (1951a). Tissue treatment. (Doctrine on biogenic stimulators). I. Background, methods and the clinical tissue treatment. *Priroda* 11, 39–46.
- Filatov, V. P. (1951b). Tissue treatment. (Doctrine on biogenic stimulators). II. Hypothesis of tissue therapy, or the doctrine on biogenic stimulators. *Priroda* 12, 20–28.
- Finnie, J. F., and van Staden, J. (1985). Effect of seaweed concentrate and applied hormones on in vitro cultured tomato roots. *J. Plant Physiol.* 120, 215–222. doi: 10.1016/S0176-1617(85)80108-5
- Fleming, C. C., Turner, S. J., and Hunt, M. (2006). Management of root knot nematodes in turfgrass using mustard formulations and biostimulants. *Commun. Agric. Appl. Biol. Sci.* 71, 653–658.
- Forde, B. G., and Lea, P. J. (2007). Glutamate in plants: metabolism, regulation, and signalling. *J. Exp. Bot.* 58, 2339–2358. doi: 10.1093/jxb/erm121
- Fuentes-Ramirez, L. E., and Caballero-Mellado, J. (2006). “Bacterial biofertilizers,” in *PGPR: Biocontrol and Biofertilization*, ed Z. A. Siddiqui (Dordrecht: Springer), 143–172.
- Gafurov, R. G., and Zefirov, N. S. (2007). A role of the molecular structure of phyto-regulators in chemical signal perception by receptors of plant hormonal systems. *Mosc. Univ. Chem. Bull.* 62, 52–56. doi: 10.3103/S0027131407010129

- Gajic, B. R. (1989). *Bioregulator Based on Plant Raw Materials and Method for Producing Same*. U.S. Patent No. 4872900. Available online at: <http://www.freepatentsonline.com/4872900.html>
- Gandarillas Infante, M. J. (2012). *Biostimulant Composition for Growth and Production of Capsicum chinense L. that Comprises a Carbohydrate Solution Originating from Chitosan and a Partial Hydrolysate of Saccharomyces Cerevisiae and Method for the Use of said Composition*. Patent CL: WO2012045189A2, 16. Available online at: <http://www.freepatentsonline.com/WO2012045189A2.html>
- García, A. C., Izquierdo, F. G., and Berbara, R. L. L. (2014). "Effects of humic materials on plant metabolism and agricultural productivity," in *Emerging Technologies and Management of Crop Stress Tolerance*, ed P. Ahmad (Amsterdam; Boston, MA; Heidelberg; London; New York, NY; Oxford; Paris; San Diego, CA; San Francisco, CA; Singapore; Sydney, NSW; Tokyo: Elsevier Inc.), 449–466.
- García, A. C., Santos, L. A., Izquierdo, F. G., Sperandio, M. V. L., Castro, R. N., and Berbara, R. L. L. (2012). Vermicompost humic acids as an ecological pathway to protect rice plant against oxidative stress. *Ecol. Eng.* 47, 203–208. doi: 10.1016/j.ecoleng.2012.06.011
- Gates, D. E., Parker, C., Boston, C. L., Allen, A., Pabon, A., Nelson, M. L., et al. (2012). "Effects of biostimulants containing fermentation metabolites and nutrients on plant response to abiotic stress," in *Abstracts Book for Oral and Poster Presentations of the 1st World Congress on the use of Biostimulants in Agriculture*; 2012 Nov 26–29, eds P. Perata, P. Brown, and M. Ponchet (Strasbourg: New Ag International), 96.
- Gawrońska, H. (2008). *Biostimulators in Modern Agriculture. General Aspects*. Warsaw: Wiesz Jutra.
- Gerhardson, B. (2002). Biological substitutes for pesticides. *Trends Biotechnol.* 20, 338–343. doi: 10.1016/S0167-7799(02)00201-8
- Ghannam, A., Abbas, A., Alek, H., Al-Waari, Z., and Al-Ktaifani, M. (2013). Enhancement of local plant immunity against tobacco mosaic virus infection after treatment with sulphated-carrageenan from red alga (*Hypnea musciformis*). *Physiol. Mol. Plant.* 84, 19–27. doi: 10.1016/j.pmp.2013.07.001
- Giannattasio, M., Vendramin, E., Fornasier, F., Alberghini, S., Zanardo, M., Stellin, F., et al. (2013). Microbiological features and bioactivity of a fermented manure product (Preparation 500) used in biodynamic agriculture. *J. Microbiol. Biotechnol.* 23, 644–651. doi: 10.4014/jmb.1212.12004
- Goatley, J. M., and Schmidt, R. E. (1991). Biostimulator enhancement of Kentucky bluegrass sod. *HortScience* 26, 254–255.
- Goni, O., Fort, A., Quille, P., McKeown, P. C., Spillane, C., and O'Connell, S. (2016). Comparative transcriptome analysis of two *Ascomycetum nodosum* extract biostimulants: same seaweed but different. *J. Agr. Food Chem.* 64, 2980–2989. doi: 10.1021/acs.jafc.6b00621
- González, A., Castro, J., Vera, J., and Moenne, A. (2013). Seaweed oligosaccharides stimulate plant growth by enhancing carbon and nitrogen assimilation, basal metabolism, and cell division. *J. Plant Growth Regul.* 32, 443–448. doi: 10.1007/s00344-012-9309-1
- González, A., Contreras, R. A., Zúiga, G., and Moenne, A. (2014). Oligo-carrageenan kappa-induced reducing redox status and activation of TRR/TRX system increase the level of indole-3-acetic acid, gibberellin A3 and trans-zeatin in *Eucalyptus globulus* trees. *Molecules* 19, 12690–12698. doi: 10.3390/molecules190812690
- Gordon, D. M. (1947). The treatment of retinitis pigmentosa with special reference to the Filatov method. *Am. J. Ophthalmol.* 30, 565–580. doi: 10.1016/0002-9394(47)92310-6
- Gupta, U. C., and MacLeod, J. A. (1982). Effect of sea crop 16 and ergostim on crop yields and plant composition. *Can. J. Soil Sci.* 62, 527–532. doi: 10.4141/cjss82-057
- Halmann, M. (1990). Synthetic plant growth regulators. *Adv. Agron.* 43, 47–105. doi: 10.1016/S0065-2113(08)60476-9
- Halpern, M., Bar-Tal, A., Ofek, M., Minz, D., Muller, T., and Yermiyahu, U. (2015). The use of biostimulants for enhancing nutrient uptake. *Adv. Agron.* 130, 141–174. doi: 10.1016/bs.agron.2014.10.001
- Hammad, S. A. R., and Ali, O. A. M. (2014). Physiological and biochemical studies on drought tolerance of wheat plants by application of amino acids and yeast extract. *Ann. Agr. Sci.* 59, 133–145. doi: 10.1016/j.aos.2014.06.018
- Hamza, B., and Suggars, A. (2001). Biostimulants: myths and realities. *TurfGrass Trends* 8, 6–10. Available online at: <http://archive.lib.msu.edu/tic/tgtre/2001.html#aug>; <http://archive.lib.msu.edu/tic/tgtre/article/2001aug6.pdf>
- Hanafy, M. S., Saadawy, F. M., Milad, S. M. N., and Ali, R. M. (2012). Effect of some natural extracts on growth and chemical constituents of *Schefflera arboricola* plants. *J. Hort. Sci. Ornament. Plants* 4, 26–33. Available online at: [http://idosi.org/jhsop/jhsop4\(1\)12.htm](http://idosi.org/jhsop/jhsop4(1)12.htm); [http://idosi.org/jhsop/4\(1\)12/4.pdf](http://idosi.org/jhsop/4(1)12/4.pdf)
- Hayes, M., García-García, M., Fitzgerald, C., and Lafarga, T. (2015). "Seaweed and milk derived bioactive peptides and small molecules in functional foods and cosmeceuticals," in *Biotechnology of Bioactive Compounds: Sources and applications*, eds V. K. Gupta and M. G. Tuohy (Chichester: John Wiley and Sons, Ltd.), 669–691.
- Henda, Y. B., and Bordenave-Juchereau, S. (2014). "Using marine cryptides against metabolic syndrome," in *Bioactive Compounds from Marine Foods: Plant and Animal Sources*, eds B. Hernández-Ledesma and M. Herrero (Chichester: John Wiley & Sons, Ltd.), 95–112.
- Herbretreau, F., Coiffard, L. J. M., Derrien, A., and De Roeck-Holtzhauer, Y. (1997). The fatty acid composition of five species of macroalgae. *Bot. Mar.* 40, 25–27.
- Hernandez, O. L., Calderín, A., Huelva, R., Martínez-Balmori, D., Guridi, F., Aguiar, N. O., et al. (2015). Humic substances from vermicompost enhance urban lettuce production. *Agron. Sustain. Dev.* 35, 225–232. doi: 10.1007/s13593-014-0221-x
- Hernandez-Herrera, R. M., Santacruz-Ruvalcaba, F., Ruiz-Lopez, M. A., Norrie, J., and Hernandez-Carmona, G. (2014). Effect of liquid seaweed extracts on growth of tomato seedlings (*Solanum lycopersicum* L.). *J. Appl. Phycol.* 26, 619–628. doi: 10.1007/s10811-013-0078-4
- Herve, J. J. (1994). Biostimulants, a new concept for the future; prospects offered by the chemistry of synthesis and biotechnology. *Comptes Rendus Acad. Agric. Fr.* 80, 91–102.
- Hirsch, R., Hartung, W., and Gimmler, H. (1989). Absciscic acid content of algae under stress. *Bot. Acta* 102, 326–334. doi: 10.1111/j.1438-8677.1989.tb00113.x
- Huang, B. (2007). Plant growth regulators: what and why. *Golf Course Management* 157–160.
- Ijdo, M., Cranenbrouck, S., and Declerck, S. (2010). Methods for large-scale production of AM fungi: past, present, and future. *Mycorrhiza* 21, 1–16. doi: 10.1007/s00572-010-0337-z
- Ikrina, M. A., and Kolbin, A. M. (2004). *Regulators of Plant Growth and Development, Vol. 1, Stimulants*. Moscow: Chimia.
- Ivanov, V. T. (2010). Peptides as universal biological regulators. *Her. Russ. Acad. Sci.* 80, 419–429. doi: 10.1134/S1019331610050011
- Janas, K. M., and Posmyk, M. M. (2013). Melatonin, an underestimated natural substance with great potential for agricultural application. *Acta Physiol. Plant* 35, 3285–3292. doi: 10.1007/s11738-013-1372-0
- Jannin, L., Arkoun, M., Etienne, P., Lainé, P., Goux, D., Garnica, M., et al. (2013). Brassica napus growth is promoted by *Ascomycetum nodosum* (L.) Le Jol. seaweed extract: microarray analysis and physiological characterization of N, C, and S metabolisms. *J. Plant Growth Regul.* 32, 31–52. doi: 10.1007/s00344-012-9273-9
- Jannin, L., Arkoun, M., Ourry, A., Lainé, P., Goux, D., Garnica, M., et al. (2012). Microarray analysis of humic acid effects on *Brassica napus* growth: involvement of N, C and S metabolisms. *Plant Soil.* 359, 297–319. doi: 10.1007/s11104-012-1191-x
- Jenkins, T. A. (2014). *Bio-Stimulant for Improved Plant Growth and Development*. U.S. Patent No. 20140162877A1, 7. Available online at: <http://www.freepatentsonline.com/y2014/0162877.html>
- Johnson, C. W. (2006). What are emergent properties and how do they affect the engineering of complex systems? *Reliability Eng. Syst. Safety* 91, 1475–1481. doi: 10.1016/j.res.2006.01.008
- Kachroo, A., and Kachroo, P. (2009). Fatty acid-derived signals in plant defense. *Ann. Rev. Phytopathol.* 47, 153–176. doi: 10.1146/annurev-phyto-080508-081820
- Karnok, K. J. (2000). Promises, promises: can biostimulants deliver? *Golf Course Manag.* 68, 67–71.
- Kauffman, G. L., Kneivel, D. P., and Watschke, T. L. (2007). Effects of a biostimulant on the heat tolerance associated with photosynthetic capacity, membrane thermostability, and polyphenol production of perennial ryegrass. *Crop Sci.* 47, 261–267. doi: 10.2135/cropsci2006.03.0171

- Kelting, M., Harris, J. R., Fanelli, J., and Appleton, B. (1998). Humate-based biostimulants affect early post-transplant root growth and sapflow of red maple. *HortScience* 33, 342–344.
- Kershengolts, B. M., Zhuravskaya, A. N., Filippova, G. V., An'shakova, V. V., Shein, A. A., Khlebnyy, E. S., et al. (2008). Innovative nano-biotechnologies in decision of problems in northern regions of Russia. *Bull. Yakut. State Univ.* 5, 20–31. Available online at: <http://cyberleninka.ru/article/n/innovatsionnye-nanobiotehnologii-v-reshenii-aktualnyh-problem-severnnyh-regionov-rossii>
- Khan, W., Hiltz, D., Critchley, A. T., and Prithiviraj, B. (2011). Bioassay to detect *Ascophyllum nodosum* extract-induced cytokinin-like activity in *Arabidopsis thaliana*. *J. Appl. Phycol.* 23, 409–414. doi: 10.1007/s10811-010-9583-x
- Khan, W., Prithiviraj, B., and Smith, D. L. (2003). Chitosan and chitin oligomers increase phenylalanine ammonia-lyase and tyrosine ammonia-lyase activities in soybean leaves. *J. Plant Physiol.* 160, 859–863. doi: 10.1078/0176-1617-00905
- Khan, W., Rayirath, U. P., Subramanian, S., Jithesh, M. N., Rayorath, P., Hodges, D. M., et al. (2009). Seaweed extracts as biostimulants of plant growth and development. *J. Plant Growth Regul.* 28, 386–399. doi: 10.1007/s00344-009-9103-x
- Kinnersley, A. M. (1993). The role of phytochelates in plant growth and productivity. *Plant Growth Regul.* 12, 207–218. doi: 10.1007/BF00027200
- Kleiber, T., and Markiewicz, B. (2013). Application of “Tytanit” in greenhouse tomato growing. *Acta Sci. Pol. Hortorum Cultus* 12, 117–126. Available online at: <http://www.acta.media.pl/pl/action/getfull.php?id=3436>
- Kolomaznik, K., Pecha, J., Friebrová, V., Janáčová, D., and Vašek, V. (2012). Diffusion of biostimulators into plant tissues. *Heat Mass Transfer.* 48, 1505–1512. doi: 10.1007/s00231-012-0998-6
- Korosov, A. V. (2012). An emergent principle in ecology. *Princ. Ėkol.* 1, 48–66. doi: 10.15393/j1.art.2012.1481
- Krouk, G. (2015). “How plants respond to a combination of signals: are biostimulant effects triggered by a cocktail of molecules?” in *Abstracts Book for Oral and Poster Presentations of the 2nd World Congress on the Use of Biostimulants in Agriculture; 2015 Nov 16–19*, eds P. Perata, P. Brown, R. A. Alvarez, and M. Ponchet (Florence: New Ag International), 28.
- Krouk, G. (2016). Hormones and nitrate: a two-way connection. *Plant Mol. Biol.* 91, 599–606. doi: 10.1007/s11103-016-0463-x
- Krouk, G., Crawford, N. M., Coruzzi, G. M., and Tsay, Y. F. (2010). Nitrate signaling: adaptation to fluctuating environments. *Curr. Opin. Plant Biol.* 13, 265–272. doi: 10.1016/j.pbi.2009.12.003
- Krouk, G., Ruffel, S., Gutiérrez, R. A., Gojon, A., Crawford, N. M., Coruzzi, G. M., et al. (2011). A framework integrating plant growth with hormones and nutrients. *Trends Plant Sci.* 16, 178–182. doi: 10.1016/j.tplants.2011.02.004
- Krouk, G., Tranchina, D., Lejay, L., Cruikshank, A. A., Shasha, D., Coruzzi, G. M., et al. (2009). A systems approach uncovers restrictions for signal interactions regulating genome-wide responses to nutritional cues in *Arabidopsis*. *PLoS Comput. Biol.* 5:e1000326. doi: 10.1371/journal.pcbi.1000326
- Kumar, D., and Shivay, Y. S. (2008). *Definitional Glossary of Agricultural Terms. V. I. IK*. New Delhi: International Publishing House Pvt Ltd.
- Kunicki, E., Grabowska, A., Sękara, A., and Wojciechowska, R. (2010). The effect of cultivar type, time of cultivation, and biostimulant treatment on the yield of spinach (*Spinacia oleracea* L.). *Folia Hort.* 22, 9–13. doi: 10.2478/fhort-2013-0153
- Kurepin, L. V., Zaman, M., and Pharis, R. P. (2014). Phytohormonal basis for the plant growth promoting action of naturally occurring biostimulators. *J. Sci. Food Agric.* 94, 1715–1722. doi: 10.1002/jsfa.6545
- Lachhab, N., Sanzani, S. M., Adrian, M., Chiltz, A., Balacey, S., Boselli, M., et al. (2014). Soybean and casein hydrolysates induce grapevine immune responses and resistance against *Plasmopara viticola*. *Front. Plant Sci.* 5:716. doi: 10.3389/fpls.2014.00716
- Leal, D., Matsuhira, B., Rossi, M., and Caruso, F. (2008). FT-IR spectra of alginic acid block fractions in three species of brown seaweeds. *Carbohydr. Res.* 343, 308–316. doi: 10.1016/j.carres.2007.10.016
- Lee, S., Yoon, J. Y., Jung, H. I., Lee, D. J., Shin, D. Y., Hyun, K. H., et al. (2012). Ameliorative effects of squash (*Cucurbita moschata* Duchesne ex Poiret) leaf extracts on oxidative stress. *Plant Growth Regul.* 67, 9–17. doi: 10.1007/s10725-011-9655-1
- Linsner, A., Cazzara, L., and Barbieri, G. (2006). Plant growth promoting rhizobacteria: a new opportunity for a sustainable agriculture. *Fertilitas Agrorum.* 1, 65–75. Available online at: http://fertilitasagrorum.ciec-italia.it/index_file/volumi.htm
- Lisiecka, J., Knaflowski, M., Spizewski, T., Fraszczak, B., Kaluzewicz, A., and Krzesinski, W. (2011). The effect of animal protein hydrolysate on quantity and quality of strawberry daughter plants cv. ‘Elsanta’. *Acta Sci. Pol.-Hortoru.* 10, 31–40. Available online at: <http://www.acta.media.pl/pl/action/getfull.php?id=2706>
- Lötze, E., and Hoffman, E. W. (2016). Nutrient composition and content of various biological active compounds of three South African-based commercial seaweed biostimulants. *J. Appl. Phycol.* 28, 1379–1386. doi: 10.1007/s10811-015-0644-z
- Lovatt, C. J. (2015). *Use of a Natural Metabolite to Increase Crop Production*. U.S. Patent Application No. 14/880,120. Available online at: <http://www.freepatentsonline.com/y2016/0088842.html>
- Lucini, L., Rouphael, Y., Cardarelli, M., Canaguier, R., Kumar, P., and Colla, G. (2015). The effect of a plant-derived biostimulant on metabolic profiling and crop performance of lettuce grown under saline conditions. *Sci. Hortic.* 182, 124–133. doi: 10.1016/j.scienta.2014.11.022
- Luisi, P. L. (2002). Emergence in chemistry: Chemistry as the embodiment of emergence. *Found. Chem.* 4, 183–200. doi: 10.1023/A:1020672005348
- Lüttge, U. (2012). Modularity and emergence: biology’s challenge in understanding life. *Plant Biol.* 14, 865–871. doi: 10.1111/j.1438-8677.2012.00659.x
- Maini, P. (2006). The experience of the first biostimulant, based on aminoacids and peptides: a short retrospective review on the laboratory researches and the practical results. *Fertilitas Agrorum.* 1, 29–43. Available online at: http://fertilitasagrorum.ciec-italia.it/index_file/volumi.htm
- Malusá, E., and Vassilev, N. A. (2014). Contribution to set a legal framework for biofertilisers. *Appl. Microbiol. Biotechnol.* 98, 6599–6607. doi: 10.1007/s00253-014-5828-y
- Malusá, E., Sas-Pasz, L., and Ciesielska, J. (2012). Technologies for beneficial microorganisms inocula used as biofertilizers. *Sci. World J.* 98, 6599–6607. doi: 10.1100/2012/491206
- Martínez-Esteso, M. J., Vilella-Antón, M. T., Sellés-Marchart, S., Martínez-Márquez, A., Botta-Català, A., Piñol-Dastis, R., et al. (2016). A DIGE proteomic analysis of wheat flag leaf treated with TERRA-SORB® foliar, a free amino acid high content biostimulant. *J. Integr. Omics* 6, 9–17. doi: 10.5584/jiomics.v6i1.188
- Martínez-Viveros, O., Jorquera, M. A., Crowley, D. E., Gajardo, G., and Mora, M. L. (2010). Mechanisms and practical considerations involved in plant growth promotion by rhizobacteria. *J. Soil Sci. Plant Nutr.* 10, 293–319. doi: 10.4067/S0718-95162010000100006
- Matsubayashi, Y., and Sakagami, Y. (2006). Peptide hormones in plants. *Annu. Rev. Plant Biol.* 57, 649–674. doi: 10.1146/annurev.arplant.56.032604.144204
- Matyjaszczyk, E. (2015). Products containing microorganisms as a tool in integrated pest management and the rules of their market placement in the European Union. *Pest Manag. Sci.* 71, 1201–1206. doi: 10.1002/ps.3986
- Mayr, E. (1982). *The Growth of Biological Thought: Diversity, Evolution, and Inheritance*. Cambridge, MA; London: Harvard University Press.
- McCarty, L. B. (2001). *Best Golf Course Management Practices*. Upper Saddle River, NJ: Prentice-Hall.
- Mercier, L., Lafitte, C., Borderies, G., Briand, X., Esquerré-Tugayé, M. T., and Fournier, J. (2001). The algal polysaccharide carrageenans can act as an elicitor of plant defence. *New Phytol.* 149, 43–51. doi: 10.1046/j.1469-8137.2001.00011.x
- Michalak, I., and Chojnacka, K. (2014). Algal extracts: Technology and advances. *Eng Life Sci.* 14, 581–591. doi: 10.1002/elsc.201400139
- Michalski, T. (2008). “Possibilities of maize production increase using non-conventional technologies,” in *Monographs series: Biostimulants in Modern Agriculture. General Aspects*, ed H. Gawrońska (Warsaw: Wiesz Jutra), 30–53.
- Migliore, M., Felici, B., Benedetti, A., and Florio, A. (2013). Proposal of bioassays as a tool for screening biostimulant properties of protein hydrolysates from animal waste materials. *Acta Hort.* 1009, 235–240. doi: 10.17660/ActaHortic.2013.1009.28
- Mikić, M., and Dobromilska, R. (2014). Assessment of yield and physiological indices of small-sized tomato cv. ‘Bianka F₁’ under the influence of biostimulators of marine algae origin. *Acta Sci. Pol.-Hortoru.* 13, 31–41. Available online at: <http://www.acta.media.pl/pl/action/getfull.php?id=3851>

- Miller, C. O., Skoog, F., Von Saltza, M. H., and Strong, F. M. (1955). Kinetin, a cell division factor from deoxyribonucleic acid. *J. Am. Chem. Soc.* 77, 1392–1392. doi: 10.1021/ja01610a105
- Mladenova, Y. I. (1978). Effect of L-glutamic acid and Siapton leaf organic fertilizer on oxidized nicotinamide adenine dinucleotide dependent glutamate dehydrogenase of different maize genotypes. *J. Agric. Food. Chem.* 26, 1274–1276. doi: 10.1021/jf60220a034
- Mladenova, Y. I., Maini, P., Mallegni, C., Goltsev, V., Vladova, R., Vinarova, K., et al. (1998). Siapton – an amino-acid-based biostimulant reducing osmotic stress metabolic changes in maize. *Agro Food Ind. Hi-Tech.* 9, 18–22.
- Mochida, K., and Shinozaki, K. (2011). Advances in omics and bioinformatics tools for systems analyses of plant functions. *Plant Cell Physiol.* 52, 2017–2038. doi: 10.1093/pcp/pcr153
- Mora, V., Bacaicoa, E., Zamarreño, A.-M., Aguirre, E., Garnica, M., Fuentes, M., et al. (2010). Action of humic acid on promotion of cucumber shoot growth involves nitrate-related changes associated with the root-to-shoot distribution of cytokinins, polyamines and mineral nutrients. *J. Plant Physiol.* 167, 633–642. doi: 10.1016/j.jplph.2009.11.018
- Murch, S. J., and Saxena, P. K. (2002). Melatonin: a potential regulator of plant growth and development? *In vitro Cell. Dev. Biol. Plant* 38, 531–536. doi: 10.1079/IVP.20020233
- Nabil, S., and Cosson, J. (1996). Seasonal variations in sterol composition of *Delesseria sanguinea* (Ceramiales, Rhodophyta). *Hydrobiologia* 326, 511–514. doi: 10.1007/BF00047854
- Nardi, S., Ertani, A., Concheri, G., and Pizzeghello, D. (2006). Metodi di determinazione dell'attività biostimolante. *Fertilitas Agrorum.* 1, 47–53. Available online at: http://fertilitasagrorum.ciec-italia.it/index_file/volumi.htm
- Nardi, S., Muscolo, A., Vaccaro, S., Baiano, S., Spaccini, R., and Piccolo, A. (2007). Relationship between molecular characteristics of soil humic fractions and glycolytic pathway and Krebs cycle in maize seedlings. *Soil Biol. Biochem.* 39, 3138–3146. doi: 10.1016/j.soilbio.2007.07.006
- Nardi, S., Pizzeghello, D., Schiavon, M., and Ertani, A. (2016). Plant biostimulants: physiological responses induced by protein hydrolyzed-based products and humic substances in plant metabolism. *Sci. Agric.* 73, 18–23. doi: 10.1590/0103-9016-2015-0006
- Nardi, S., Tosoni, M., Pizzeghello, D., Provenzano, M. R., Cilenti, A., Sturaro, A., et al. (2005). Chemical characteristics and biological activity of organic substances extracted from soils by root exudates. *Soil Sci. Soc. Am. J.* 69, 2012–2019. doi: 10.2136/sssaj2004.0401
- Naumov, G. F., Bozhkov, A. I., Leontovich, V. P., Sklyar, A. I., and Belous, A. M. (1993). Polyfunctionality of allelopathic substance allelostim. *Doklady Akademii nauk Ukrainy* 11, 166–169.
- Nelson, W. R., and van Staden, J. (1985). 1-Aminocyclopropane-1-carboxylic acid in seaweed concentrate. *Bot. Mar.* 28, 415–418.
- Okazaki, Y., and Saito, K. (2014). Roles of lipids as signaling molecules and mitigators during stress response in plants. *Plant J.* 79, 584–596. doi: 10.1111/tpj.12556
- Omar, H., Abdullatif, B., Al-Kazan, M., and El-Gendy, A. (2015). Various applications of seaweed improves growth and biochemical constituents of *Zea mays* L. and *Helianthus annuus* L. *J. Plant Nutr.* 38, 28–40. doi: 10.1080/01904167.2014.911893
- Onatsky, N. M., Yakhin, I. A., Rybalkin, S. P., Mikhina, L. V., Yakhin, O. I., and Ibatullina, R. B. (2001). Toxicological assessment of the preparation Stifun and substantiation of its MAC in the occupational air. *Toxicol. Rev.* 5, 20–24. Available online at: <http://en.toxreview.ru/>
- Painter, T. J. (1983). "Algal polysaccharides," in *The Polysaccharides*, ed G. O. Aspinall (New York, NY: Academic Press), 195–285.
- Paradičković, N., Vinković, T., Vinković Vrček, I., and Tkalec, M. (2013). Natural biostimulants reduce the incidence of BER in sweet yellow pepper plants (*Capsicum annuum* L.). *Agric. Food Sci.* 22, 307–317. Available online at: <http://ojs.tsv.fi/index.php/AFS/article/view/7354>
- Paradičković, N., Vinković, T., Vinković Vrček, I., Žuntar, I., Bojić, M., and Medić-Šarić, M. (2011). Effect of natural biostimulants on yield and nutritional quality: an example of sweet yellow pepper (*Capsicum annuum* L.) plants. *J. Sci. Food Agric.* 91, 2146–2152. doi: 10.1002/jsfa.4431
- Pardo-García, A. I., Martínez-Gil, A. M., Cadahía, E., Pardo, F., Alonso, G. L., and Salinas, M. R. (2014). Oak extract application to grapevines as a plant biostimulant to increase wine polyphenols. *Food Res. Int.* 55, 150–160. doi: 10.1016/j.foodres.2013.11.004
- Parrado, J., Bautista, J., Romero, E. J., García-Martínez, A. M., Friaiza, V., and Tejada, M. (2008). Production of a carob enzymatic extract: potential use as a biofertilizer. *Bioresour. Technol.* 99, 2312–2318. doi: 10.1016/j.biortech.2007.05.029
- Parrado, J., Escudero-Gilete, M. L., Friaiza, V., García-Martínez, A., González-Miret, M. L., Bautista, J. D., et al. (2007). Enzymatic vegetable extract with bio-active components: influence of fertiliser on the colour and anthocyanins of red grapes. *J. Sci. Food. Agric.* 87, 2310–2318. doi: 10.1002/jsfa.2989
- Pecha, J., Fürst, T., Kolomazník, K., Friebrová, V., and Svoboda, P. (2012). Protein biostimulant foliar uptake modeling: the impact of climatic conditions. *AIChE J.* 58, 2010–2019. doi: 10.1002/aic.12739
- Petrozza, A., Santaniello, A., Summerer, S., Di Tommaso, G., Di Tommaso, D., Paparelli, E., et al. (2014). Physiological responses to Megafof® treatments in tomato plants under drought stress: a phenomic and molecular approach. *Sci. Hortic.* 174, 185–192. doi: 10.1016/j.scienta.2014.05.023
- Pizzeghello, D., Francioso, O., Ertani, A., Muscolo, A., and Nardi, S. (2013). Isopentenyladenosine and cytokinin-like activity of different humic substances. *J. Geochem. Explor.* 129, 70–75. doi: 10.1016/j.gexplo.2012.10.007
- Pretorius, J. C. (2007). *Seed Suspensions from "Lupinus albus", Isolated Compounds Thereof and Use as Biological Plant Strengthening Agent*. Patent No. WO2007090438 A1, 59. Available online at: <http://www.freepatentsonline.com/WO2007090438A1.html>
- Pretorius, J. C. (2013). *Extracts and Compounds from "Agapanthus africanus" and Their Use as Biological Plant Protecting Agents*. Patent No. WO2007003286 A2. Available online at: <http://www.freepatentsonline.com/WO2007003286A2.html>
- Przybyls, A., Gawrońska, H., and Gajc-Wolska, J. (2014). Biological mode of action of a nitrophenolates-based biostimulant: case study. *Front. Plant Sci.* 5:713c. doi: 10.3389/fpls.2014.00713
- Radkowsky, A., and Radkowska, I. (2013). Effect of foliar application of growth biostimulant on quality and nutritive value of meadow sward. *Ecol. Chem. Eng. A.* 20, 1205–1211. doi: 10.2428/ceca.2013.20(10)110
- Raldugin, V. A. (2004). Triterpenoids of fir and highly efficient plant growth regulator based on them. *Russ. Chem. J.* 48, 84–88.
- Rathore, S. S., Chaudhary, D. R., Boricha, G. N., Ghosh, A., Bhatt, B. P., Zodape, S. T., et al. (2009). Effect of seaweed extract on the growth, yield and nutrient uptake of soybean (*Glycine max*) under rainfed conditions. *South Afr. J. Bot.* 75, 351–355. doi: 10.1016/j.sajb.2008.10.009
- Rattan, R. S. (2010). Mechanism of action of insecticidal secondary metabolites of plant origin. *Crop Prot.* 29, 913–920. doi: 10.1016/j.cropro.2010.05.008
- Ravensberg, W. J. (2015). "Commercialisation of microbes: present situation and future prospects," in *Principles of Plant-Microbe Interactions*, ed B. Lugtenberg (Cham; Heidelberg; New York, NY; Dordrecht; London: Springer International Publishing), 309–317.
- Rayorath, P., Khan, W., Palanisamy, R., MacKinnon, S. L., Stefanova, R., Hankins, S. D., et al. (2008). Extracts of the brown seaweed *Ascophyllum nodosum* induce gibberellic acid (GA3)-independent amylase activity in barley. *J. Plant Growth Regul.* 27, 370–379. doi: 10.1007/s00344-008-9063-6
- Rengasamy, K. R. R., Kulkarni, M. G., Stirk, W. A., and Van Staden, J. (2015a). Eckol improves growth, enzyme activities, and secondary metabolite content in maize (*Zea mays* cv. Border King). *J. Plant Growth Regul.* 34, 410–416. doi: 10.1007/s00344-015-9479-8
- Rengasamy, K. R. R., Kulkarni, M. G., Stirk, W. A., and Van Staden, J. (2015b). Eckol - a new plant growth stimulant from the brown seaweed *Ecklonia maxima*. *J. Appl. Phycol.* 27, 581–587. doi: 10.1007/s10811-014-0337-z
- Rivera, C. M., Salerno, A., Sequi, P., Rea, E., and Trinchera, A. (2010). Exploring biostimulant effect of a brassicacea plant extract: use of maize seedling development as reference bioassay. *Acta Hortic.* 884, 737–744. doi: 10.17660/ActaHortic.2010.884.100
- Rodríguez-Morgado, B., Gómez, I., Parrado, J., and Tejada, M. (2014). Behaviour of oxyfluorfen in soils amended with edaphic biostimulants/biofertilizers obtained from sewage sludge and chicken feathers. Effects on soil biological properties. *Environ. Sci. Pollut. R.* 21, 11027–11035. doi: 10.1007/s11356-014-3040-3
- Rose, M. T., Patti, A. F., Little, K. R., Brown, A. L., Jackson, W. R., and Cavagnaro, T. R. (2014). A meta-analysis and review of plant-growth response to humic

- substances: practical implications for agriculture. *Adv. Agron.* 124, 37–89. doi: 10.1016/B978-0-12-800138-7.00002-4
- Rouse, R. E. (1984). Evaluation of two commercially available biostimulants on citrus. *J. Rio Grande Valley Hortic. Soc.* 37, 107–112.
- Roy, R. N., Finck, A., Blair, G. J., and Tandon, H. L. S. (2006). *Plant Nutrition for Food Security-A Guide for Integrated Nutrient Management*. FAO Fertilizer and Plant Nutrition Bulletin 16, Rome: FAO.
- Ruiz, J. M., Castilla, N., and Romero, L. (2000). Nitrogen metabolism in pepper plants applied with different bioregulators. *J. Agric. Food Chem.* 48, 2925–2929. doi: 10.1021/jf990394h
- Russo, R. O., and Berlyn, G. P. (1991). The use of organic biostimulants to help low-input sustainable agriculture. *J. Sustain. Agric.* 1, 19–42. doi: 10.1300/J064v01n02_04
- Saa, S., Olivos-Del Rio, A., Castro, S., and Brown, P. H. (2015). Foliar application of microbial and plant based biostimulants increases growth and potassium uptake in almond (*Prunus dulcis* Mill. D. A. Webb). *Front. Plant Sci.* 6:87. doi: 10.3389/fpls.2015.00087
- Samir, P., and Link, A. J. (2011). Analyzing the cryptome: uncovering secret sequences. *AAPS J.* 13, 152–158. doi: 10.1208/s12248-011-9252-2
- Sánchez-Gómez, R., Zalacain, A., Alonso, G. L., and Salinas, M. R. (2014). Vine-shoot waste aqueous extracts for re-use in agriculture obtained by different extraction techniques: phenolic, volatile, and mineral compounds. *J. Agric. Food Chem.* 62, 10861–10872. doi: 10.1021/jf503929v
- Sanders, D. C., Ricotta, J. A., and Hodges, L. (1990). Improvement of carrot stands with plant biostimulants and fluid drilling. *HortScience* 25, 181–183.
- Sanderson, K. J., and Jameson, P. E. (1986). The cytokinins in a liquid seaweed extract: could they be the active ingredients? *Acta Hortic.* 179, 113–116.
- Santaniello, A., Di Tommaso, D., Povero, D., Piaggese, A., and Perata, P. (2015). “Comparative analysis of the effects induced by three different *Ascophyllum nodosum* extracts: Algae® Fert BASE, SOLID and SOLID G,” in *Abstracts Book for Oral and Poster Presentations of the 2st World Congress on the use of Biostimulants in Agriculture*; 2015 Nov 16–19, eds P. Perata, P. Brown, R. A. Alvarez, and M. Ponchet (Florence: New Ag International), 100.
- Santaniello, A., Giorgi, F. M., Di Tommaso, D., Di Tommaso, G., Piaggese, A., and Perata, P. (2013). Genomic approaches to unveil the physiological pathways activated in Arabidopsis treated with plant-derived raw extracts. *Acta Hortic.* 1009, 161–174. doi: 10.17660/ActaHortic.2013.1009.20
- Satish, L., Rameshkumar, R., Rathinapriya, P., Pandian, S., Rency, A. S., Sunitha, T., et al. (2015). Effect of seaweed liquid extracts and plant growth regulators on *in vitro* mass propagation of brinjal (*Solanum melongena* L.) through hypocotyl and leaf disc explants. *J. Appl. Phycol.* 27, 993–1002. doi: 10.1007/s10811-014-0375-6
- Schiavon, M., Ertani, A., and Nardi, S. (2008). Effects of an alfalfa protein hydrolysate on the gene expression and activity of enzymes of the tricarboxylic acid (TCA) Cycle and nitrogen metabolism in *Zea mays* L. *J. Agric. Food Chem.* 56, 11800–11808. doi: 10.1021/jf802362g
- Schiavon, M., Pizzeghello, D., Muscolo, A., Vaccaro, S., Francioso, O., and Nardi, S. (2010). High molecular size humic substances enhance phenylpropanoid metabolism in maize (*Zea mays* L.). *J. Chem. Ecol.* 36, 662–669. doi: 10.1007/s10886-010-9790-6
- Schmidt, R. E. (1992). Biostimulants. *Grounds Maintenance* 1992, 27, 38–56.
- Schmidt, R. E., Ervin, E. H., and Zhang, G. (2003). Questions and answers about biostimulants. *Golf Course Management* 71, 91–94.
- Sharma, H. S. S., Fleming, C., Selby, C., Rao, J. R., and Martin, T. (2014). Plant biostimulants: a review on the processing of macroalgae and use of extracts for crop management to reduce abiotic and biotic stresses. *J. Appl. Phycol.* 26, 465–490. doi: 10.1007/s10811-013-0101-9
- Sharma, K., Bruns, C., Butz, A. F., and Finckh, M. R. (2012). Effects of fertilizers and plant strengtheners on the susceptibility of tomatoes to single and mixed isolates of *Phytophthora infestans*. *Eur. J. Plant Pathol.* 133, 739–751. doi: 10.1007/s10658-012-9954-z
- Sharma, S. H. S., Lyons, G., McRoberts, C., McCall, D., Carmichael, E., Andrews, F., et al. (2012a). Biostimulant activity of brown seaweed species from Strangford Lough: compositional analyses of polysaccharides and bioassay of extracts using mung bean (*Vigna mungo* L.) and pak choi (*Brassica rapa chinensis* L.). *J. Appl. Phycol.* 24, 1081–1091. doi: 10.1007/s10811-011-9737-5
- Sharma, S. H. S., Lyons, G., McRoberts, C., McCall, D., Carmichael, E., Andrews, F., et al. (2012b). Brown seaweed species from Strangford Lough: compositional analyses of seaweed species and biostimulant formulations by rapid instrumental methods. *J. Appl. Phycol.* 24, 1141–1157. doi: 10.1007/s10811-011-9744-6
- Sharp, R. G. (2013). A Review of the applications of chitin and its derivatives in agriculture to modify plant-microbial interactions and improve crop yields. *Agronomy* 3, 757–793. doi: 10.3390/agronomy3040757
- Sleighter, R. L., Caricasole, P., Richards, K. M., Hanson, T., and Hatcher, P. G. (2015). “Molecular level characterization of humic substances and correlation with plant growth stimulation,” in *Abstracts Book for Oral and Poster Presentations of the 2st World Congress on the use of Biostimulants in Agriculture 2015 Nov 16–19*, eds P. Perata, P. Brown, R. A. Alvarez, and M. Ponchet (Florence: New Ag International), 10.
- Smeekeens, S. (2000). Sugar-induced signal transduction in plants. *Annu. Rev. Plant Phys.* 51, 49–81. doi: 10.1146/annurev.arplant.51.1.49
- Smoleń S. (2012). “Foliar nutrition: current state of knowledge and opportunities,” in *Advances in citrus nutrition*, ed A. K. Srivastava (Dordrecht; Heidelberg; New York, NY; London: Springer Science+Business Media BV), 41–58.
- Sofo, A., Nuzzaci, M., Vitti, A., Tataranni, G., and Scopa, A. (2014). “Control of biotic and abiotic stresses in cultivated plants by the use of biostimulant microorganisms,” in *Improvement of Crops in the Era of Climatic Changes*, eds P. Ahmad, M. R. Wani, M. M. Azooz, and L. S. Phan Tran (New York, NY: Springer Science+Business Media), 107–117.
- Spaepen, S. (2015). “Plant hormones produced by microbes,” in *Principles of plant-microbe interactions*, ed B. Lugtenberg (Cham; Heidelberg; New York, NY; Dordrecht; London: Springer International Publishing), 247–256.
- Spinelli, F., Fiori, G., Noferini, M., Sprocati, M., and Costa, G. (2010). A novel type of seaweed extract as a natural alternative to the use of iron chelates in strawberry production. *Sci. Hortic.* 125, 263–269. doi: 10.1016/j.scienta.2010.03.011
- Srivastava, A., Bhatia, G., Pant, R., and Srivastava, P. C. (2010). Bioefficacy and residue studies of Fantac (biostimulant) in rice crop under sub-tropical conditions. *J. Environ. Prot.* 1, 261–263. doi: 10.4236/jep.2010.13031
- Srivastava, A., Bhatia, G., and Srivastava, P. C. (2008). Persistence behavior of Fantac biostimulant in chili and soil under subtropical conditions. *B. Environ. Contam. Tox.* 80, 403–406. doi: 10.1007/s00128-008-9390-0
- Stadnik, M. J., and de Freitas, M. B. (2014). Algal polysaccharides as source of plant resistance inducers. *Trop. Plant Pathol.* 39, 111–118. doi: 10.1590/S1982-56762014000200001
- Stirk, W. A., and Staden, J. (1996). Comparison of cytokinin- and auxin-like activity in some commercially used seaweed extracts. *J. Appl. Phycol.* 8, 503–508. doi: 10.1007/BF02186328
- Stirk, W. A., and Van Staden, J. (2006). “Seaweed products as biostimulants in agriculture,” in *World Seaweed Resources*, eds A. T. Critchley, M. Ohno, and D. B. Largo (Amsterdam: ETI Information Services Ltd.), 1–32.
- Stirk, W. A., Novák, O., Strnad, M., and Van Staden, J. (2003). Cytokinins in macroalgae. *Plant Growth Regul.* 41, 13–24. doi: 10.1023/A:1027376507197
- Stirk, W. A., Tarkowská, D., Turečková, V., Strnad, M., and van Staden, J. (2014). Absciscic acid, gibberellins and brassinosteroids in Kelpak®, a commercial seaweed extract made from *Ecklonia maxima*. *J. Appl. Phycol.* 26, 561–567. doi: 10.1007/s10811-013-0062-z
- Sukhoverkhov, F. M. (1967). “The effect of cobalt, vitamins, tissue preparations and antibiotics on carp production,” in *Proceedings of the FAO world Symposium on Warm-Water Pond Fish Culture* (Rome), 44, 400–407.
- Summerer, S., Petrozza, A., and Cellini, F. (2013). High throughput plant phenotyping: a new and objective method to detect and analyse the biostimulant properties of different products. *Acta Hortic.* 1009, 143–148. doi: 10.17660/ActaHortic.2013.1009.17
- Tachibana, S., Summer, P., Ewing, J., Miwa, T., and Kitazawa, D. (2012). *Method of Producing Plant Biostimulant*. U.S. Patent No. 20120129695 A1 p. 14. Available online at: <http://www.freepatentsonline.com/y2012/0129695.html>
- Tagliavini, S., and Kubiskin, C. (2006). Effetti della biostimolazione in ortofrutticoltura: alcune esperienze a confronto. *Fertilitas Agrorum* 1, 23–28. Available online at: http://fertilitasagrorum.ciec-italia.it/index_file/volumi.htm
- Tandon, S., and Dubey, A. (2015). Effects of Biozyme (*Ascophyllum nodosum*) biostimulant on growth and development of soybean [*Glycine max* (L.) Merill]. *Commun. Soil Sci. Plant Anal.* 46, 845–858. doi: 10.1080/00103624.2015.1011749

- Tay, S. A. B., Palni, L. M. S., and MacLeod, J. K. (1987). Identification of cytokinin glucosides in seaweed extract. *J. Plant Growth Regul.* 5, 133–138. doi: 10.1007/BF02087181
- Tegeder, M. (2012). Transporters for amino acids in plant cells: some functions and many unknowns. *Curr. Opin. Plant. Biol.* 15, 315–321. doi: 10.1016/j.pbi.2012.02.001
- Tejada, M., Benitez, C., and Parrado, J. (2011a). Application of biostimulants in benzo(a)pyrene polluted soils: short-time effects on soil biochemical properties. *Appl. Soil Ecol.* 50, 21–26. doi: 10.1016/j.apsoil.2011.08.002
- Tejada, M., Benitez, C., Gymeza, I., and Parrado, J. (2011b). Use of biostimulants on soil restoration: effects on soil biochemical properties and microbial community. *Appl. Soil Ecol.* 49, 11–17. doi: 10.1016/j.apsoil.2011.07.009
- Tejada, M., García-Martínez, A. M., Gómez, I., and Parrado, J. (2010). Application of MCPA herbicide on soils amended with biostimulants: short-time effects on soil biological properties. *Chemosphere* 80, 1088–1094. doi: 10.1016/j.chemosphere.2010.04.074
- Tejada, M., Rodríguez-Morgado, B., Gómez, I., and Parrado, J. (2014). Degradation of chlorpyrifos using different biostimulants/biofertilizers: effects on soil biochemical properties and microbial community. *Appl. Soil Ecol.* 84, 158–165. doi: 10.1016/j.apsoil.2014.07.007
- Thao, H. T. B., and Yamakawa, T. (2009). Phosphite (phosphorous acid): fungicide, fertilizer or bio-stimulator? *Soil Sci. Plant. Nutr.* 55, 228–234. doi: 10.1111/j.1747-0765.2009.00365.x
- Thomas, M., Chauhan, D., Patel, J., and Panchal, T. (2013). Analysis of biostimulants made by fermentation of *Sargassum tenerimum* seaweed. *Int. J. Curr. Trop. Res.* 2, 405–407.
- Tian, S. K., Lu, L. L., Xie, R. H., Zhang, M. Z., Jernstedt, J. A., Hou, D. D., et al. (2015). Supplemental macronutrients and microbial fermentation products improve the uptake and transport of foliar applied zinc in sunflower (*Helianthus annuus* L.) plants. Studies utilizing micro X-ray fluorescence. *Front. Plant Sci.* 6:87. doi: 10.3389/fpls.2014.00808
- Torre, L. A., Battaglia, V., and Caradonia, F. (2013). Legal aspects of the use of plant strengtheners (biostimulants) in Europe. *Bulg. J. Agric. Sci.* 19, 1183–1189. Available online at: <http://www.agrojournal.org/19/06-02.pdf>; <http://www.agrojournal.org/19/06-02.html>
- Torre, L. A., Battaglia, V., and Caradonia, F. (2016). An overview of the current plant biostimulant legislations in different European Member States. *J. Sci. Food Agric.* 96, 727–734. doi: 10.1002/jsfa.7358
- Traon, D., Amat, L., Zotz, F., and du Jardin, P. (2014). *A Legal Framework for Plant Biostimulants and Agronomic Fertiliser Additives in the EU*. Report to the European Commission, DG Enterprise & Industry, Arcadia International, 115.
- Trevisan, S., Francioso, O., Quaggiotti, S., and Nardi, S. (2010). Humic substances biological activity at the plant-soil interface from environmental aspects to molecular factors. *Plant Signal. Behav.* 5, 635–643. doi: 10.4161/psb.5.6.11211
- Turan, M., and Köse, C. (2004). Seaweed extracts improve copper uptake of grapevine. *Acta Agric. Scand. B* 54, 213–220. doi: 10.1080/09064710410030311
- Ugolini, L., Cinti, S., Righetti, L., Stefan, A., Matteo, R., D'Avino, L., et al. (2015). Production of an enzymatic protein hydrolyzate from defatted sunflower seed meal for potential application as a plant biostimulant. *Ind. Crop Prod.* 75, 15–23. doi: 10.1016/j.indcrop.2014.11.026
- Ulrich-Merzenich, G., Panek, D., Zeitler, H., Wagner, H., and Vetter, H. (2009). New perspectives for synergy research with the “omic”-technologies. *Phytomedicine* 16, 495–508. doi: 10.1016/j.phymed.2009.04.001
- Valepyn, E., Cabrera, J. C., Richel, A., and Paquot, M. (2014). Water soluble exo-polysaccharide from *Syncephalastrum racemosum*, a strong inducer of plant defence reactions. *Carbohydr Polym.* 101, 941–946. doi: 10.1016/j.carbpol.2013.10.018
- Van der Watt, E., and Pretorius, J. C. (2011). *In vitro* and *in vivo* bio-stimulatory properties of a *Lupinus albus* L. seed suspension. *Crop Pasture Sci.* 62, 189–197. doi: 10.1071/CP10391
- Van der Watt, E., and Pretorius, J. C. (2013). A triglyceride from *Lupinus albus* L. seed with biostimulatory properties. *Afr. J. Biotechnol.* 35, 5431–5443. doi: 10.5897/AJB12.2851
- Van Regenmortel, M. H. (2004). Reductionism and complexity in molecular biology. *EMBO Rep.* 5, 1016–1020. doi: 10.1038/sj.embor.7400284
- Vasconcelos, A. C. F., Zhang, X., Ervin, E. H., and Kiehl, J. C. (2009). Enzymatic antioxidant responses to biostimulants in maize and soybean subjected to drought. *Sci. Agric.* 66, 395–402. doi: 10.1590/S0103-90162009000300015
- Vaskova, H., Kolomaznik, K., and Vasek, V. (2013). Hydrolysis process of collagen protein from tannery waste materials for production of biostimulator and its mathematical model. *Int. J. Math. Mod. Methods Appl. Sci.* 7, 568–575. Available online at: <http://naun.org/cms.action?id=5358>
- Vera, J., Castro, J., Gonzalez, A., and Moenne, A. (2011). Seaweed polysaccharides and derived oligosaccharides stimulate defense responses and protection against pathogens in plants. *Mar. Drugs* 9, 2514–2525. doi: 10.3390/md9122514
- Verkleij, F. N. (1992). Seaweed extracts in agriculture and horticulture: a review. *Biol. Agric. Hortic.* 8, 309–324. doi: 10.1080/01448765.1992.9754608
- Vernieri, P., Borghesi, E., Ferrante, A., and Magnani, G. (2005). Application of biostimulants in floating system for improving rocket quality. *J. Food Agric. Environ.* 3, 86–88. Available online at: <http://world-food.net/application-of-biostimulants-in-floating-system-for-improving-rocket-quality/>
- Vernieri, P., Borghesi, E., Toggoni, F., Serra, G., Ferrante, A., and Piagessi, A. (2006). Use of biostimulants for reducing nutrient solution concentration in floating system. *Acta Hortic.* 718, 477–484. doi: 10.17660/ActaHortic.2006.718.55
- Vesela, M., and Friedrich, J. (2009). Amino acid and soluble protein cocktail from waste keratin hydrolysed by a fungal keratinase of *Paecilomyces marquandii*. *Biotechnol. Bioprocess Eng.* 14, 84–90. doi: 10.1007/s12257-008-0083-7
- Vessey, J. K. (2003). Plant growth promoting rhizobacteria as biofertilizers. *Plant Soil.* 255, 571–586. doi: 10.1023/A:1026037216893
- Vijayanand, N., Ramya, S. S., and Rathinavel, S. (2014). Potential of liquid extracts of *Sargassum wightii* on growth, biochemical and yield parameters of cluster bean plant. *Asian Pac. J. Reprod.* 3, 150–155. doi: 10.1016/S2305-0500(14)60019-1
- Vinoth, S., Gurusaravanan, P., and Jayabalan, N. (2014). Optimization of somatic embryogenesis protocol in *Lycopersicon esculentum* L. using plant growth regulators and seaweed extracts. *J. Appl. Phycol.* 26, 1527–1537. doi: 10.1007/s10811-013-0151-z
- Viriji, M. G. (2007). *A Plant Derived Biostimulant Formulation and Method for Preparation Thereof*. Patent No. WO2007052282 A1. Available online at: <http://www.freepatentsonline.com/WO2007052282A1.html>
- Vyas, S., Guha, S., Bhattacharya, M., and Rao, I. U. (2009). Rapid regeneration of plants of *Dendrobium lituiflorum* Lindl. (Orchidaceae) by using banana extract. *Sci Hortic.* 121, 32–37. doi: 10.1016/j.scienta.2009.01.012
- Wang, Y. H., and Irving, H. R. (2011). Developing a model of plant hormone interactions. *Plant Signal. Behav.* 6, 494–500. doi: 10.4161/psb.6.4.14558
- Weissbach, H., King, W., Sjoerdsma, A., and Udenfriend, S. (1959). Formation of indole-3-acetic acid and tryptamine in animals: a method for estimation of indole-3-acetic acid in tissue. *J. Biol. Chem.* 234, 81–86.
- Wilson, H. T., Xu, K., and Taylor, A. G. (2015). Transcriptome analysis of gelatin seed treatment as a biostimulant on cucumber plant growth. *ScientificWorldJournal* 2015:391234. doi: 10.1155/2015/391234
- Wintermute, E. H., and Silver, P. A. (2010). Emergent cooperation in microbial metabolism. *Mol. Syst. Biol.* 6:407. doi: 10.1038/msb.2010.66
- Wu, A. (2016). *Hot Bio-Stimulants Gain Traction in Latin American market*. Available online at: <http://news.agropages.com/News/NewsDetail--19117.htm> (Accessed August 25, 2016).
- Wu, Y., Jenkins, T., Blunden, G., von Mende, N., and Hankins, S. D. (1998). Suppression of fecundity of the root-knot nematode, *Meloidogyne javanica*, in monoxenic cultures of *Arabidopsis thaliana* treated with an alkaline extract of *Ascophyllum nodosum*. *J. Appl. Phycol.* 10, 91–94. doi: 10.1023/A:1008067420092
- Xavier, I. J., and Boyetchko, S. M. (2002). “Arbuscular mycorrhizal fungi as biostimulants and bioprotectants of crops,” in *Applied Mycology and Biotechnology. Agriculture and Food Production*, Vol. 2, eds G. G. Khachatourians and D. K. Arora (Amsterdam: Elsevier), 311–330.
- Yakhin, I. A., Ibragimov, R. I., Yakhin, O. I., Isaev, R. F., and Vakhitov, V. A. (1998). The induced effect of biopreparation stifun on the accumulation of trypsin inhibitors in potato tubers during storage. *Russ. Agric. Sci.* 4, 12–13.
- Yakhin, O. I., Lubyantov, A. A., and Yakhin, I. A. (2012). Changes in cytokinins, auxin, and abscisic acid contents in wheat seedlings treated with the growth regulator stifun. *Russ. J. Plant Physiol.* 59, 398–405. doi: 10.1134/S1021443712030193

- Yakhin, O. I., Lubyantov, A. A., Yakhin, I. A., and Vakhitov, V. A. (2007). Protective role of bioregulator stifun under the negative effect of cadmium. *Russ. Agric. Sci.* 33, 233–235. doi: 10.3103/S1068367407040064
- Yakhin, O. I., Lubyantov, A. A., Yakhin, I. A., Vakhitov, V. A., Ibragimov, R. I., Yumaguzhin, M. S., et al. (2011a). Metabolic changes in wheat (*Triticum aestivum* L.) plants under action of bioregulator stifun. *Appl. Biochem. Micro.* 47, 621–626. doi: 10.1134/S0003683811060123
- Yakhin, O. I., Lubyantov, A. A., and Yakhin, I. A. (2014). Modern concepts on biostimulators. *Agrokhimiya* 7, 85–90. Available online at: <http://elibrary.ru/item.asp?id=21779483>
- Yakhin, O. I., Lubyantov, A. A., and Yakhin, I. A. (2016a). Biostimulants in agrotechnologies: problems, solutions, outlook. *Agrochemical Her.* 1, 15–21. Available online at: <http://www.agrochemv.ru/en/nomer/2016/1>; <http://elibrary.ru/item.asp?id=25940647>
- Yakhin, O. I., Lubyantov, A. A., and Yakhin, I. A. (2016b). Physiological activity and efficiency of application of biostimulants. *Agrokhimiya* 6, 72–94. Available online at: <http://elibrary.ru/item.asp?id=26470157>
- Yakhin, O. I., Lubyantov, A. A., Yakhin, I. A., Gareeva, G. B., Markelova, E. M., Kabirov, R. R., et al. (2013). Ecological evaluation of plant growth regulator stifun with the use of a multi-component test system. *Agrokhimiya* 3, 65–71. Available online at: <http://elibrary.ru/item.asp?id=18962629>
- Yakhin, O. I., Lubyantov, A. A., Yakhin, I. A., Postrikan, B. N., Chemeris, A. V., Vakhitov, V. A., et al. (2011b). Effect of the plant growth regulator stifun on the accumulation of cadmium in grain crops. *Agrokhimiya* 5, 76–83. Available online at: <http://elibrary.ru/item.asp?id=16356500>
- Yakhin, O. I., Yakhin, I. A., Spirikhin, L. V., and Khalilov, L. M. (2005). “Development of methods for standardization of multicomponent bioregulators based on vegetable raw materials,” in *Mass Spectrometry and Its Application Problems. II Congress RSMS. Russian Conference with International Participation* (Moscow), MBS-19.
- Yakhin, O. I., Yakhin, I. A., Vakhitov, V. A., and Lubyantov, A. A. (2006). The mechanism of action of the natural bioregulator stifun. *Dokl. Biochem Biophys.* 411, 327–330. doi: 10.1134/S1607672906060019
- Yamaguchi, Y., and Huffaker, A. (2011). Endogenous peptide elicitors in higher plants. *Curr. Opin. Plant Biol.* 14, 351–357. doi: 10.1016/j.pbi.2011.05.001
- Yasmeen, A., Basra, S. M. A., Farooq, M., Rehman, H., Hussain, N., and Athar, H. R. (2013). Exogenous application of moringa leaf extract modulates the antioxidant enzyme system to improve wheat performance under saline conditions. *Plant Growth Regul.* 69, 225–233. doi: 10.1007/s10725-012-9764-5
- Yasmeen, A., Nouman, W., Basra, S. M. A., Wahid, A., Rehman, H., Hussain, N., et al. (2014). Morphological and physiological response of tomato (*Solanum lycopersicum* L.) to natural and synthetic cytokinin sources: a comparative study. *Acta Physiol. Plant.* 36, 3147–3155. doi: 10.1007/s11738-014-1662-1
- Zandonadi, D. B., Canellas, L. P., and Façanha, A. R. (2007). Indolacetic and humic acids induce lateral root development through a concerted plasmalemma and tonoplast H⁺ pumps activation. *Planta* 225, 1583–1595. doi: 10.1007/s00425-006-0454-2
- Zandonadi, D. B., Santos, M. P., Dobbss, L. B., Olivares, F. L., Canellas, L. P., Binzel, M. L., et al. (2010). Nitric oxide mediates humic acids-induced root development and plasma membrane H⁺-ATPase activation. *Planta* 231, 1025–1036. doi: 10.1007/s00425-010-1106-0
- Zhang, X., and Schmidt, R. (1999). Biostimulating turfgrasses. *Grounds Maintenance* 34, 14–15.
- Zhang, X., and Schmidt, R. E. (2000). Hormone containing products impact on antioxidant status of tall fescue and creeping bent grass subjected to drought. *Crop Sci.* 40, 1344–1349. doi: 10.2135/cropsci2000.4051344x
- Zhao, J., Davis, L. C., and Verpoorte, R. (2005). Elicitor signal transduction leading to production of plant secondary metabolites. *Biotechnol. Adv.* 23, 283–333. doi: 10.1016/j.biotechadv.2005.01.003
- Ziosi, V., Zandoli, R., Di Nardo, A., Biondi, S., Antognoni, F., and Calandriello, F. (2013). Biological activity of different botanical extracts as evaluated by means of an array of *in vitro* and *in vivo* bioassays. *Acta Hort.* 1009, 69–66. doi: 10.17660/ActaHortic.2013.1009.5
- Zodape, S. T. (2001). Seaweeds as a biofertilizer. *J. Sci. Ind. Res. India.* 60, 378–382. Available online at: <http://nopr.niscair.res.in/handle/123456789/26485>
- Zodape, S. T., Gupta, A., Bhandari, S. C., Rawat, U. S., Cahudhary, D. R., Eswara, K., et al. (2011). Foliar application of seaweed sap as biostimulant for enhancement of yield and yield quality of tomato (*Lycopersicon esculentum* Mill.). *J. Sci. Ind. Res. India.* 70, 215–219. Available online at: <http://nopr.niscair.res.in/handle/123456789/11089>

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 Yakhin, Lubyantov, Yakhin and Brown. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.